



UNIVERSIDADE FEDERAL DE SERGIPE
CENTRO DE CIÊNCIAS BIOLÓGICAS E DA SAÚDE
DEPARTAMENTO DE MEDICINA

**EPIDEMIOLOGIA MOLECULAR DE GASTROENTERITE
AGUDA CAUSADA POR NOROVIRUS EM CRIANÇAS
MENORES DE 5 ANOS A NÍVEL MUNDIAL: REVISÃO
SISTEMÁTICA E META-ANÁLISE**

HIRAM MENEZES NASCIMENTO FILHO

Aracaju-SE

2018

**EPIDEMIOLOGIA MOLECULAR DE GASTROENTERITE
AGUDA CAUSADA POR NOROVIRUS EM CRIANÇAS
MENORES DE 5 ANOS A NÍVEL MUNDIAL: REVISÃO
SISTEMÁTICA E META-ANÁLISE**

Monografia apresentada ao Colegiado do Curso de Medicina da Universidade Federal de Sergipe como requisito parcial para conclusão da graduação em Medicina.

Acadêmico: Hiram Menezes Nascimento Filho

Orientador: Prof. Dr. Ricardo Queiroz Gurgel

Aracaju-SE

2018

**Epidemiologia molecular de gastroenterite aguda causada por Norovírus em
crianças menores de 5 anos: Revisão Sistemática e Meta-Análise.**

Monografia apresentada à Universidade Federal de Sergipe como requisito parcial à conclusão da graduação de Medicina do Centro de Ciências Biológicas e da Saúde.

Autor: Hiram Menezes Nascimento Filho

Orientador: Prof. Dr. Ricardo Queiroz Gurgel

**Epidemiologia molecular de gastroenterite aguda causada por Norovírus em
crianças menores de 5 anos: Revisão Sistemática e Meta-Análise.**

Monografia apresentada à Universidade Federal de Sergipe como requisito parcial à conclusão da graduação de Medicina do Centro de Ciências Biológicas e da Saúde.

Aprovada em: ____ de _____ de _____

BANCA EXAMINADORA

Universidade Federal de Sergipe

Universidade Federal de Sergipe

Universidade Federal de Sergipe

AGRADECIMENTOS

Agradeço a Deus por guiar o meu caminho, permitir o aprendizado e colocar na minha vida pessoas que irradiam luz.

Aos meus pais, Hiram e Bernadete, por me fazer um filho honrado em tê-los como pais, por me instruir no caminho da educação, da ética e da perseverança.

A minha irmã, Hayla, e minha namorada, Luisa, pelo incentivo em prol das minhas conquistas, pela mão estendida no momento de dificuldade e por mostrar que sonhar é a fonte de energia que move nossas conquistas.

Aos professores da Universidade Federal de Sergipe, médicos do Hospital Universitário, e todos os funcionários que dedicaram parte do seu trabalho a compartilhar comigo e meus colegas seus conhecimentos técnicos e experiências.

Ao Prof. Ricardo Gurgel que ao longo da graduação despertou minha crítica frente ao conhecimento científico, norteou minha prática em função da medicina baseada em evidências e ensinou que a arte caminha de mãos dadas com a ciência médica.

*“A esperança tem duas filhas lindas, a indignação e
a coragem: a indignação nos ensina a não aceitar
as coisas como estão; a coragem a mudá-las”.*

(Santo Agostinho)

LISTA DE ABREVIATURAS E SIGLAS

ADN	Adenovírus
AGVI	Aliança Global para Vacinações e Imunizações
EIA	Ensaio Imunoenzimático
ELISA	Ensaio Imunoabsorvente mediado por Enzimas
GEA	Gastroenterite Aguda
IgA	Imunoglobulina A
IgG	Imunoglobulina G
MS	Ministério da Saúde
ODM	Objetivo de Desenvolvimento do Milênio
OMS	Organização Mundial da Saúde
PCR	Reação em Cadeia da Polimerase
ROV	Rotavírus
SBP	Sociedade Brasileira de Pediatria
SF 0,9%	Soro Fisiológico concentrado a 0,9%
SRO	Soro de Reidratação Oral
VLP	Partícula semelhante a Vírus

LISTA DE TABELAS

Table 1- Norovirus specimens by genogroups from 2006 to 2015.....65

LISTA DE FIGURAS

Figura 1- Nutrição, saúde e ambiente têm grande importância na prevenção e tratamento da gastroenterite aguda infantil	13
Figura 2- Filogenia de Norovírus, baseado no sequenciamento de aminoácidos da proteína VP1.....	17
Figure 1- Flow diagram of study selection.....	64
Figure 2- Norovirus circulating genotypes from 2006 to 2008.....	66
Figure 3- Norovirus circulating genotypes from 2009 to 2011.....	67
Figure 4- Norovirus circulating genotypes from 2006 to 2008.....	68

SUMÁRIO

1. REVISÃO DE LITERATURA	11
1.1. GASTROENTERITE AGUDA	11
1.1.1 Impacto em Saúde.....	11
1.1.2 Definição e Etiopatogenia.....	12
1.1.3 Fatores de Risco	13
1.1.4 Determinação da Severidade da Doença.....	13
1.1.5 Conduta Terapêutica	14
1.2. NOROVÍRUS	15
1.2.1. Histórico.....	15
1.2.2. Características e Evolução	16
1.2.3. Diagnóstico Laboratorial	18
1.2.4. Prevenção e Controle	19
2. REFERÊNCIAS BIBLIOGRÁFICAS.....	21
3. NORMAS PARA PUBLICAÇÃO.....	36
3.1. JOURNAL INFORMATION	36
3.2. SCOPE	36
3.3. SYSTEMATIC REVIEWS AND META-ANALYSES	36
3.4. CRITERIA FOR PUBLICATION.....	37
3.5. STYLE AND FORMAT	41
4. ARTIGO ORIGINAL.....	45
4.1. ABSTRACT.....	46
4.2. INTRODUCTION	47
4.3. METHODS	48
4.4. RESULTS	50
4.5. DISCUSSION	53
4.6. REFERENCES	54
4.7. SUPPORTING INFORMATION	63

1 REVISÃO DE LITERATURA

1.1. GASTROENTERITE AGUDA

1.1.1 Impacto em Saúde

A gastroenterite aguda (GEA) revela-se importante causa de morbidade e mortalidade em crianças no mundo. Estima-se que essa patologia cause 526.000 mortes em menores de 5 anos anualmente, sendo a 4^a principal causa de mortalidade (Liu *et al.*, 2016). O combate à GEA compreende, portanto, parte fundamental do Objetivo de Desenvolvimento do Milênio (ODM) de redução da mortalidade infantil, afirmado pela Declaração do Milênio das Nações Unidas (United Nations, 2000).

Anualmente os episódios ambulatoriais de GEA respondem por aproximadamente U\$325 milhões em custos médicos diretos e U\$423 milhões em custos à sociedade. Observam-se, entretanto, dois cenários distintos entre os países desenvolvidos e os países em desenvolvimento. Aqueles enfrentam as consequências decorrentes de hospitalizações, dias perdidos de produtividade e afastamento dos responsáveis das atividades laborais. Enquanto isso os últimos concentram alta carga da doença, lutando contra a mortalidade provocada pela mesma (Rheingans *et al.*, 2009).

Em virtude do conhecimento de que o Rotavírus (ROV) era o principal agente etiológico responsável por morbidade e mortalidade por GEA em crianças menores de 5 anos, em 2009 a Organização Mundial de Saúde (OMS) recomendou a inclusão da vacina contra ROV nos programas nacionais de imunização dos países onde a mortalidade por GEA representava valores maiores ou iguais a 10% nessa faixa etária mesmo se a eficácia avaliada não fosse elevada (WHO, 2009).

O Brasil tornou-se pioneiro na vacinação contra ROV, introduzindo a vacina monovalente Rotarix® no seu Programa Nacional de Imunização em 2006. O país alcançou elevados índices de vacinação na população infantil (85%), e assim como Estados Unidos da América, Finlândia, Bélgica, El Salvador, México, Panamá e Nicarágua atingiu redução na incidência de GEA por RoV, bem como hospitalizações e mortalidade por gastroenterite de todas as causas (Gurgel *et al.*, 2009; Orozco *et al.*, 2009; Richardson *et al.*, 2010; Sáfadi *et al.*, 2010).

Mesmo com o incentivo proporcionado pela Aliança em Vacina (GAVI Global Alliance for Vaccines and Immunization), a vacinação contra ROV é dispendiosa para o sistema de saúde, especialmente para países de média e baixa renda. Somado a isso, sabe-se que os indicadores de redução na mortalidade infantil exigem medidas mais amplas, saneamento, tratamento de água, oferta de atenção básica à saúde (Loganathan *et al.*, 2016).

1.1.2 Definição e Etiopatogenia

Define-se GEA a diminuição da consistência das fezes (amolecidas ou líquidas) e/ou frequência maior ou igual a três vezes por dia, com ou sem vômitos, de duração inferior a 14 dias (Guarino *et al.*, 2014).

Essencialmente a GEA é estabelecida por 5 mecanismos etiopatogênicos: osmótico, secretor, invasivo, por redução da motilidade, por redução da área de superfície intestinal. A desidratação é a principal complicação da GEA, sendo a faixa etária menor de 5 anos o grupo mais vulnerável aos distúrbios hidroeletrólíticos e acidobásicos provocados pela doença (Pediatria, 2014).

Entre os casos de gastroenterite em crianças menores de 5 anos, identificam-se patógenos entéricos na maioria das amostras de fezes. Os vírus são os principais agentes implicados na etiopatogenia da doença (70%), liderados por ROV e NOV, enquanto as bactérias *Escherichia coli*, *Campylobacter jejuni*, *Yersinia* spp, *Shigella* spp, *Salmonella* spp, *Clostridium difficile*, e protozoários, *Cryptosporidium parvum*, *Giardia intestinalis*, *Entamoeba histolytica* contam com papel de significância no diagnóstico diferencial (Elliott, 2007).

O ROV, que por muitos anos foi alvo de estudos na temática da GEA, é responsável pelos quadros mais severos. Atua por mecanismos osmótico e secretor através da toxina NSP4 (Halaihel *et al.*, 2000). O NOV, também conhecido como Vírus de Norwalk, ainda não possui mecanismo bem estabelecido. Aceita-se o postulado de apoptose de enterócitos do intestino proximal, entretanto não se sabe se por efeito viral direto ou toxina (Troeger *et al.*, 2008).

Desde o licenciamento das vacinas contra ROV, o NOV emergiu como principal agente etiológico de GEA em crianças menores de 5 anos em países que atingiram altos índices de cobertura vacinal. Embora se trate de agente responsável por doença de alto impacto em saúde, provoca quadro clínico mais brando que o ROV (Bartsch *et al.*, 2016; Hemming *et al.*, 2013; Lanata *et al.*, 2013; Lopman *et al.*, 2016; Orozco *et al.*, 2009; Riera-Montes, O’Ryan e Verstraeten, 2017).

1.1.3 Fatores de Risco

Enquanto o agente etiológico tem papel significante na evolução da GEA, existem fatores de risco inerentes ao hospedeiro associados à severidade e persistência da doença: (1) Idade - Lactentes menores de 6 meses são mais suscetíveis a episódios diarreicos persistentes e severos (Strand *et al.*, 2012). (2) Alimentação - Crianças alimentadas com amamentação exclusiva durante os primeiros seis meses de vida e que são amamentadas até os dois anos apresentam menos infecções e quadros patológicos menos graves (Victora e Barros, 2000). (3) Status imunológico - Imunodeficiência e doenças crônicas são fatores de risco para diarréia por agentes oportunistas como *Clostridium difficile* e *Cryptosporidium* (Vecchio, Lo e Zucur, 2012). (4) Condições Socioeconômicas – Baixa cobertura sanitária e acesso limitado ao sistema de saúde estão associados a maior mortalidade por GEA (Kotloff *et al.*, 2013). Figura 1.

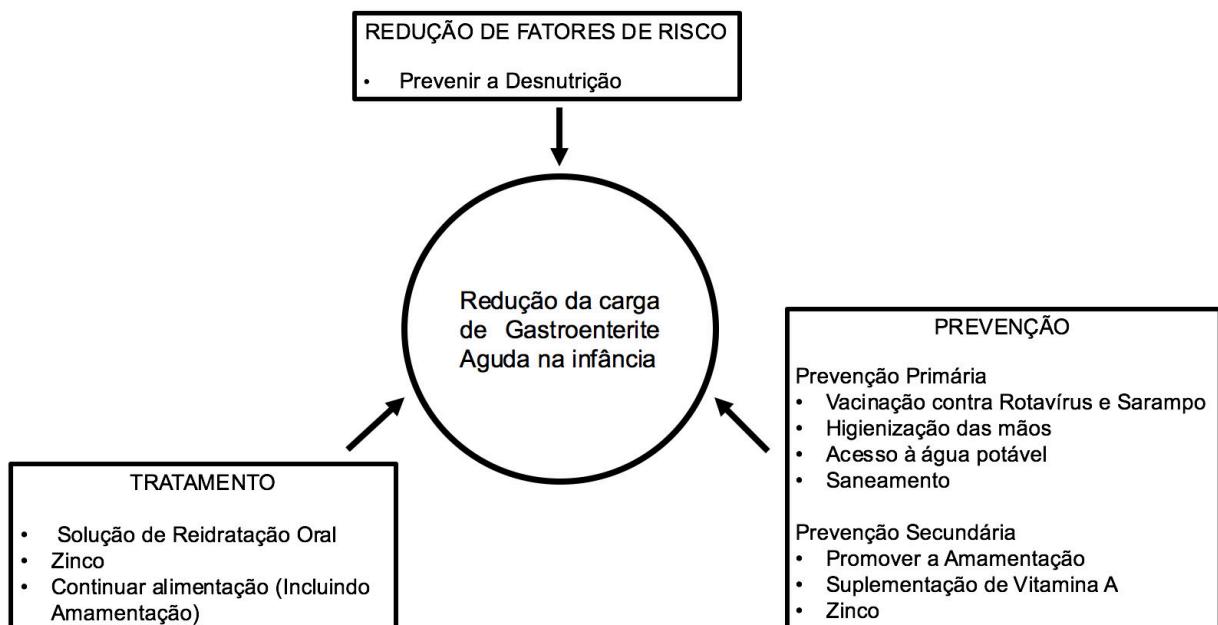


Figura 1. Adaptado de Grupo de Referência Saúde Epidemiológica da Criança, 2009
 Nutrição, saúde e ambiente têm grande importância na prevenção e tratamento da Gastroenterite Aguda infantil

1.1.4 Determinação da Severidade da Doença

As estratégias de tratamento da GEA baseiam-se essencialmente na determinação do grau de desidratação da criança. Durante os ensaios clínicos das vacinas contra ROV foram desenvolvidas as escalas de Vesikari e Clark. Amplamente difundidas no meio científico, avaliam número de episódios e duração de diarreia e vômitos, bem como a temperatura do paciente. Embora forneçam subsídio para as decisões clínicas, não atestam o estado nutricional, estado de alerta, diurese e capacidade de ingerir líquidos (Fired Clark *et al.*, 1988; Ruuska e Vesikari, 1990).

O Ministério da Saúde (MS) e a Sociedade Brasileira de Pediatria (SBP) endossam a orientação da OMS na avaliação da criança com GEA. O estado de hidratação do paciente com diarreia deve ser criteriosamente avaliado por meio de exame do estado geral, dos olhos, quanto a presença de lágrimas, se o paciente tem sede, nível de consciência, sinal da prega cutânea, pulso e tempo de enchimento capilar (Ministério da Saúde, 2012; WHO, 2005; Rodrigues, 2017).

1.1.5 Conduta Terapêutica

Embora não exista evidência científica de forte impacto, recomenda-se fortemente a internação hospitalar de crianças que se apresentam com choque, desidratação severa, alteração neurológica, vômitos incoercíveis, falha na reidratação oral, suspeita de patologia cirúrgica ou quaisquer outras condições cujo manejo ambulatorial não é seguro (Guarino *et al.*, 2014).

A hidratação é o padrão-ouro para o tratamento da GEA, contando com o Soro de Rehidratação Oral (SRO) como grande pilar da terapêutica (Hartling *et al.*, 2006). O MS define três planos terapêuticos que guiam a via de administração e volume de reposição volêmica de acordo com o grau de desidratação do paciente. O Plano A, aplicado em pacientes hidratados, orienta o aumento da ingesta hídrica de modo a prevenir desidratação. Mantém-se a alimentação habitual e instrui-se quanto aos sinais de alarme. O Plano B, aplicado em pacientes desidratados, consiste na administração do SRO. O paciente segue sob observação e avaliação do estado de hidratação. O Plano C, aplicado em pacientes francamente desidratados e desidratados refratários ao uso de SRO, consiste na administração de salina isotônica via parenteral em uma fase rápida e uma fase de manutenção. Uma vez capaz de ingerir líquido pela via oral, será iniciado SRO concomitante à reposição intravenosa (Ministério da Saúde, 2012).

Os pacientes submetidos ao Plano B, quando refratários à desidratação, podem se beneficiar de reposição volêmica por via nasogástrica. Sabe-se que a gastróclise está associada a mínimos

eventos adversos e permanência hospitalar mais breve que a via parenteral, entretanto não é comumente indicada pelos profissionais de saúde no manejo da GEA (Freedman *et al.*, 2011).

Uma vez iniciado o Plano C, a fase de expansão deverá ser executada com Salina Isotônica (SF 0,9%). Esta constituição diminui o risco de hiponatremia e suas graves consequências (Neville *et al.*, 2006). Reestabelecida a volemia, segue-se com a administração de solução acrescida de glicose e cloreto de potássio para a manutenção da necessidade basal do paciente.

No caso de pacientes que se apresentam com sangue nas fezes e queda do estado geral deve ser iniciada antibioticoterapia empírica com uso de Quinolona ou Betalactâmico com cobertura para bactérias Gram negativas (Ministério da Saúde, 2012).

No tocante às medidas adjuvantes no tratamento da GEA, investe-se na administração de zinco suplementar, alimentação livre de lactose e uso de probióticos.

Revisão de literatura conduzida em 2013 identificou importante efeito da suplementação com zinco na redução da duração de diarreia em crianças entre 6 meses e 5 anos. Os principais beneficiados com as medidas foram crianças residentes em países em desenvolvimento, onde ainda se registra significante deficiência do nutriente (Lazzerini e Ronfani, 2013).

Embora registre importância no manejo de uma consequência patológica, o consumo de produtos livres de lactose está associado à redução na duração da diarreia em crianças hospitalizadas. A agressão à mucosa intestinal pelos agentes causadores da doença reduz a expressão de enzimas responsáveis pela degradação do carboidrato, contribuindo com a manutenção das perdas digestivas (MacGillivray, Fahey e McGuire, 2013).

Muito se discute quanto ao uso de moduladores da microbiota intestinal no tratamento da gastroenterite aguda. Cepas de *Lactobacillus rhamnosus* e *Saccharomyces boulardii* podem ser consideradas no manejo da doença associadas ao SRO. Considerando-se a pequena força de evidência dessa medida e o alto custo inerente, recomenda-se avaliar a relação custo/benefício (Vandenplas *et al.*, 2012).

1.2. NOROVÍRUS

1.2.1. Histórico

Data de 1929, por Zahorsky, a primeira descrição de uma gastroenterite epidêmica não causada por bactérias. Foi denominada naquela oportunidade “Winter Vomiting Disease” e caracterizada por início súbito de náusea, vômitos, diarreia e febre baixa, envolvendo estudantes e indivíduos institucionalizados, com disseminação secundária para familiares. Em 1969, na cidade de Norwalk, Ohio, 50% dos estudantes e professores de uma escola de ensino fundamental desenvolveram quadro clínico semelhante ao descrito por Zahorsky em intervalo de 24 horas. Nas 48 horas seguintes identificou-se taxa de ataque secundário de 33%, e ao longo das semanas seguintes novos casos foram notificados, mantendo os potenciais padrões de transmissão pessoa-pessoa e fonte comum. No decorrer da investigação amostras de fezes de 39 indivíduos foram avaliadas para a presença de *Salmonella* sp., *Shigella* sp., *Escherichia coli* enteropatogênica, *Staphylococcus aureus* e vibriões não coléricos, sem sucesso. Propusera-se a hipótese de que um agente viral era responsável pela gênese da doença (Adler e Zickl, 1969).

1.2.2. Características e Evolução

O NOV é conhecido na literatura por seu formato pequeno e circular, bem como sua similaridade com o agente de Norwalk. Trata-se de um vírus de estrutura icosaédrica, cujo genoma é formado por fita única de RNA, de tamanho de 7.7kb, envolvido por um capsídeo proteico (Proteína VP1) sem envelope e com depressões características na superfície. Visualmente lembra um cálice, sendo classificado por taxonomia na família *Caliciviridae* (Glass, Parashar e Estes, 2009).

O NOV registra grande diversidade (Figura 2). Dentre os 6 genogrupos existentes, 3 (GI, GII e GIV) estão implicados nas infecções em seres humanos. As cepas virais são classificadas de acordo com o genótipo, determinado pela sequência genética codificadora da proteína VP1 (Zheng *et al.*, 2006).

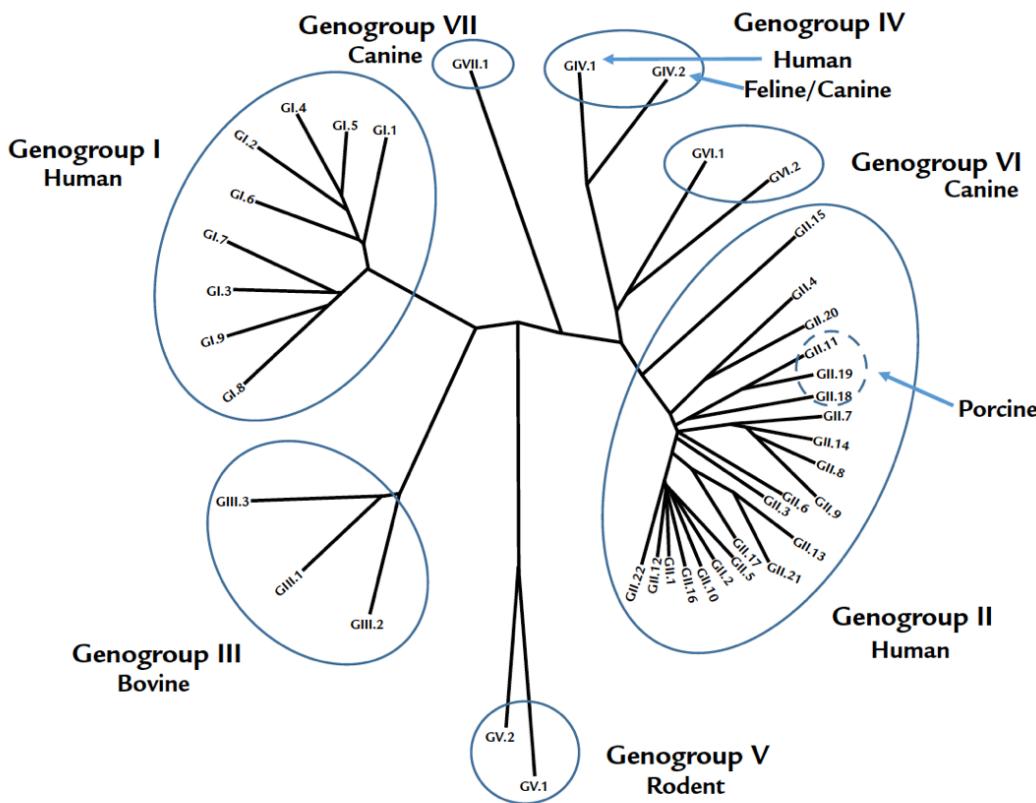


Figura 2. Adaptado de N.W. Cortes-Penfield et al.

Filogenia de Norovírus, baseado no sequenciamento dos aminoácidos da proteína VP1

Pelo menos 40 genótipos já foram isolados de amostras de fezes de indivíduos com GEA. Destaque para a cepa designada GII.4 (Genogruppo II, genótipo 4), responsável por aproximadamente 80% das infecções por NOV (Siebenga, J. Joukje *et al.*, 2009).

Desde a década de 1990 observa-se que algum genótipo predomina mundialmente nos eventos infecciosos por NOV. Estima-se que a cada 2-3 anos uma nova variante assuma o protagonismo. GII.4 US95/96 (1996), seguido de GII.4 Farmington Hills (2002), GII.4 Hunter (2004), GII.4 Den Haag (2006b), GII.4 New Orleans (2009), GII.4 Sydney (2012), foram identificadas como as variantes predominantes nos respectivos anos com quebra desse domínio pelo GII.4 em 2015 em Houzhou, China, quando foi identificado o GII.17 como grande responsável por surtos de GEA locais, depois de registro do mesmo genótipo na Austrália (Beek, van *et al.*, 2013; Bull *et al.*, 2006; Eden *et al.*, 2010; Fankhauser *et al.*, 1998; Han *et al.*, 2015; Lopman *et al.*, 2004; Yen *et al.*, 2011).

Acredita-se que o comportamento evolutivo do NOV se assemelha ao Vírus Influenza. Esse comportamento pode explicar a contínua evasão aos mecanismos de defesa dos hospedeiros (Bull e White, 2011). Análise Bayesiana usando somente dados de NOV isolados no território da China propuseram uma taxa de $4.74 \text{ a } 7.68 \times 10^{-3}$ substituições de nucleotídeos por território e por ano entre 1978 e 2000 (Qiao, Wang e Liu, 2016).

1.2.3. Diagnóstico Laboratorial

Em um passado recente, as técnicas de Microscopia Eletrônica direta e imune eram aplicadas em amostras de fezes para o diagnóstico de NOV. Apesar de especificidade considerável, a sensibilidade do teste era dependente da coleta de amostras por até 3 dias após o início do quadro clínico, bem como do treinamento do operador da técnica (Schmid *et al.*, 2004).

Atualmente identifica-se crescente implementação de métodos moleculares na rotina diagnóstica dos laboratórios. As técnicas de inoculação viral em culturas de tecidos, incentivadas pelo Programa Global de Erradicação de Poliovírus, vêm sendo substituídas pelo sequenciamento do gene codificador da proteína VP1(She *et al.*, 2010). Contando com elevadas taxas de sensibilidade e especificidade, os métodos diagnósticos derivados da Reação em Cadeia da Enzima Polimerase (PCR) mantêm-se como padrão-ouro para diagnóstico de GEA por NOV em amostras de fezes. Mesmo com baixa disponibilidade de carga viral nas amostras de fezes a detecção do agente costuma ser bem-sucedida (Schmid *et al.*, 2004).

Kroneman e colaboradores (2011) propuseram uma ferramenta para sequenciamento/definição de genótipo automatizada de NOV e Enterovírus pertencentes às famílias *Picornaviridae* e *Caliciviridae*. Essa ferramenta amplamente difundida pretere valores de corte para sequências de aminoácidos em prol de similaridades filogenéticas entre as cepas virais. Tal abordagem revela-se mais robusta, permitindo maior sensibilidade na identificação viral, muitas vezes dificultada pelo acúmulo de mutações ao longo do tempo (Kroneman *et al.*, 2011).

Finalmente, as técnicas de ensaio imunológico (EIA, ELISA) revelam papel limitado no diagnóstico de NOV. Embora facilmente conduzidas, apresentam limitações diagnósticas por conta da alta diversidade genética e antigênica do agente (Bruin, de *et al.*, 2006). Quando dois kits comerciais de ensaios imunoenzimáticos, IDEIA® e Ridascreen® foram testados frente ao padrão-ouro (PCR), identificou-se que a sensibilidade dos testes está intimamente relacionada ao padrão

de comportamento das infecções, surto de larga escala ou caso isolado. Taxas de sensibilidade mais elevadas eram alcançadas quando 6 ou mais amostras de um surto eram avaliadas. Recomenda-se, portanto, que os testes imunoenzimáticos podem servir para avaliação imediata, entretanto os resultados devem ser confirmados por PCR (Gray *et al.*, 2007).

1.2.4. Prevenção e Controle

Grande parte das medidas de prevenção e controle de infecção pelo NOV são direcionadas aos surtos de GEA provocados pelo agente. Embora se trate de uma doença benigna, crianças e idosos são suscetíveis a experimentar desfechos negativos, queda do estado geral, internação, terapêutica invasiva, decorrentes das complicações por GEA.

Em se tratando de ambiente hospitalar, recomenda-se o isolamento de contato por pelo menos 48h do início do quadro clínico. No caso de crianças < 2 anos, 5 dias de isolamento pode ser prudente. Além disso a higienização das mãos entre pacientes, acompanhantes e profissionais de saúde é fundamental para quebrar a cadeia de transmissão do agente. Devido à possibilidade do agente se depositar em superfícies, recomenda-se a limpeza e desinfecção de superfícies tocadas pelo potencial infectado. Certamente, a atitude mais importante é a comunicação do potencial surto aos órgãos competentes (Lee, Kuntz e Stevenson, 2013).

É reconhecido o potencial valor econômico na prevenção contra o NOV (Bartsch *et al.*, 2012). Somado à crescente prevalência do agente, surge o interesse na produção de uma vacina, campo em que 5 grupos distintos trabalham atualmente. Duas vacinas encontram-se em ensaio clínico: A primeira à base de Partículas semelhantes a Vírus (VLPs) e desenvolvida por Takeda vaccines; A segunda à base de Adenovírus recombinante expressando VP1 e desenvolvida por Vaxart (Cortes-Penfield *et al.*, 2017).

A vacina composta por VLPs utiliza componentes de NOV GI.3 e GII.4, variantes extremamente significantes do ponto de vista epidemiológico e molecular. Em estudo prévio de Fase II conduzido entre adultos saudáveis entre 18 e 49 anos, foram relatados mínimos eventos adversos sistêmicos entre os participantes com predomínio no grupo placebo quando comparado com outros dois grupos que receberam a vacina (23.5% x 15.2% x 21.4%). No tocante à resposta imunológica, ao final de 28 dias foi evidenciado aumento total de Imunoglobulinas e IgA nos dois grupos que receberam vacina, sendo a resposta contra VLPs de GII.4 mais intensa que contra VLPs

GI.3 (Atmar *et al.*, 2016). Entre 2012 e 2013 a mesma vacina foi testada em termos de eficácia por meio de ensaio clínico controlado e duplo-cego em busca de comparar os desfechos clínicos dos participantes. Dentre os 98 participantes (50 vacinados e 48 não vacinados) 57 investigados desenvolveram GEA por NOV quando inoculados com uma cepa desafio (54.0% dos vacinados x 62.5% dos não vacinados). Embora a incidência da doença tenha sido semelhante, ao comparar a história da doença entre os participantes a vacina mostrou-se protetora contra formas graves da doença (Bernstein *et al.*, 2015).

A vacina composta por cepa de Adenovírus (ADN) recombinante trabalha através da expressão de VP1 pertencente a variante GI.1 de NOV. Recentemente foram publicados os resultados obtidos na Fase I, que contou com 66 indivíduos alocados aleatoriamente em três grupos: Placebo, baixa dose de vacina, alta dose de vacina (20x23x23). A vacina mostrou perfil de boa tolerância e a maior parte dos eventos adversos relatados foram classificados como leves. Destaque para diarreia no grupo que recebeu alta dose de vacina, que apresentou diferença estatística confiável quando comparado ao grupo placebo ($P=0.0275$). Em relação à resposta imunológica, identificou-se elevação nos títulos de IgG e IgA contra VP1 de NoV nos grupos que receberam as vacinas, bem como persistência de resposta de IgA fecal após 6 meses da vacina (Kim *et al.*, 2018)

2. REFERÊNCIAS BIBLIOGRÁFICAS

- ABUGALIA, M.; CUEVAS, L.; KIRBY, A.; DOVE, W.; NAKAGOMI, O.; NAKAGOMI, T.; KARA, M.; GWEDER, R.; SMEO, M. Clinical Features and Molecular Epidemiology of Rotavirus and Norovirus Infections in Libyan Children. **Journal of medical virology**, v. 83, p. 1849–1856, 2011.
- ADLER, J. L.; ZICKL, R. Winter Vomiting Disease Stable. **The Journal of Infectious Diseases**, v. 119, n. 6, p. 668–673, 1969.
- AL-RASHIDI, A.; CHEHADED, W.; SZUCS, G.; ALBERT, M. J. Different Norovirus Genotypes in Patients With Gastroenteritis in Kuwait. **Journal of Medical Virology**, v. 85, p. 1161–1618, 2013.
- ALAM, A.; QURESHI, S.; VINJÉ, J.; ZAIDI, A. Genetic Characterization of Norovirus Strains in Hospitalized Children From Pakistan. **Journal of medical virology**, v. 88, p. 216–223, 2016.
- ATMAR, R. L. *et al.* Rapid Responses to 2 Virus-Like Particle Norovirus Vaccine Candidate Formulations in Healthy Adults: A Randomized Controlled Trial. **Journal of Infectious Diseases**, v. 214, n. 6, p. 845–853, 2016.
- BARTSCH, S. M.; LOPMAN, B. A.; HALL, A. J.; PARASHAR, U. D.; LEE, B. Y. The potential economic value of a human norovirus vaccine for the United States. **Vaccine**, v. 30, n. 49, p. 7097–7104, 2012.
- BARTSCH, S. M.; LOPMAN, B. A.; OZAWA, S.; HALL, A. J.; LEE, B. Y. Global economic burden of norovirus gastroenteritis. **PLoS ONE**, v. 11, n. 4, p. 1–16, 2016.
- BECKER-DREPS, S. *et al.* Etiology of Childhood Diarrhea Following Rotavirus Vaccine Introduction: A Prospective, Population-Based Study in Nicaragua. **The Pediatric Infectious Disease Journal**, v. 33, n. 11, p. 1156–1163, 2014.

BEEK, J. VAN *et al.* Indications for worldwide increased norovirus activity associated with emergence of a new variant of Genotype II.4, LATE 2012. **Eurosurveillance**, v. 18, n. 1, p. 3–4, 2013.

BERNSTEIN, D. I. *et al.* Norovirus vaccine against experimental human GII.4 virus illness: A challenge study in healthy adults. **Journal of Infectious Diseases**, v. 211, n. 6, p. 870–878, 2015.

BODHIDATTA, L.; ABENTE, E.; NEESANANT, P.; NAKJARUNG, K.; SIRICHOTE, P.; BUNYARAKYOTHIN, G.; VITHAYASAI, N.; MASON, C. J. Molecular Epidemiology and Genotype Distribution of Noroviruses in Children in Thailand From 2004 to 2010: A Multi-Site Study. **Journal of medical virology**, v. 87, p. 664–674, 2015.

BRUIN, E. DE; DUIZER, E.; VENNEMA, H.; KOOPMANS, M. P. G. Diagnosis of Norovirus outbreaks by commercial ELISA or RT-PCR. **Journal of Virological Methods**, v. 137, n. 2, p. 259–264, 2006.

BUCARDO, F.; REYES, Y.; SVENSSON, L.; NORDGREN, J. Predominance of norovirus and sapovirus in nicaragua after implementation of universal rotavirus vaccination. **PLoS ONE**, v. 9, n. 5, p. 1–8, 2014.

BULL, R. A.; TU, E. T. V; MCIVER, C. J.; RAWLINSON, W. D.; WHITE, P. A. Emergence of a New Norovirus Genotype II . 4 Variant Associated with Global Outbreaks of Gastroenteritis. **Journal of clinical microbiology**, v. 44, n. 2, p. 327–333, 2006.

BULL, R. A.; WHITE, P. A. Mechanisms of GII.4 norovirus evolution. **Trends in Microbiology**, v. 19, n. 5, p. 233–240, 2011.

CHAIMONGKOL, N.; KHAMRIN, P.; MALASAO, R.; THONGPRACHUM, A.; KONGSRICHAPOERN, T.; UKARAPOL, N.; USHIJIMA, H.; MANEEKARN, N. Molecular Characterization of Norovirus Variants and Genetic Diversity of Noroviruses and Sapoviruses in

Thailand. **Journal of medical virology**, v. 86, p. 1210–1218, 2014.

CHAN, M. C. W.; LEUNG, T. F.; KWOK, A. K.; LEE, N.; CHAN, P. K. S. Characteristics of patients infected with norovirus GII.4 Sydney 2012, Hong Kong, China. **Emerging infectious diseases**, v. 20, n. 4, p. 658–61, abr. 2014.

CHEN, S. Y.; FENG, Y.; CHAO, H. C.; LAI, M. W.; HUANG, W. L.; LIN, C. Y.; TSAI, C. N.; CHEN, C. L.; CHIU, C. H. Emergence in Taiwan of novel norovirus GII.4 variants causing acute gastroenteritis and intestinal haemorrhage in children. **Journal of Medical Microbiology**, v. 64, n. 2015, p. 544–550, 2015.

CHEN, S. Y.; TSAI, C. N.; CHEN, C. L.; CHAO, H. C.; LEE, Y. S.; LAI, M. W.; CHEN, C. C.; HUANG, W. L.; CHIU, C. H. Severe viral gastroenteritis in children after suboptimal rotavirus immunization in Taiwan. **Pediatric Infectious Disease Journal**, v. 32, n. 12, p. 1335–1339, 2013.

CHENG, W. XIA; YE, X. HUA; YANG, X. MEI; LI, Y. NING; JIN, M.; JIN, Y.; DUAN, Z. JUN. Epidemiological study of human calicivirus infection in children with gastroenteritis in Lanzhou from 2001 to 2007. **Archives of Virology**, v. 155, n. 4, p. 553–555, 2010.

CHHABRA, P. *et al.* Viral gastroenteritis in rotavirus negative hospitalized children <5 years of age from the independent states of the former Soviet Union. **Infection, Genetics and Evolution**, v. 28, p. 283–288, 2014.

CHO, H. G.; LEE, S. G.; KIM, J. E.; YU, K. S.; LEE, D. Y.; PARK, P. H.; YOON, M. HYE; JHO, E. H.; KIM, J.; PAIK, S. Y. Molecular epidemiology of norovirus GII.4 variants in children under 5 years with sporadic acute gastroenteritis in South Korea during 2006-2013. **Journal of Clinical Virology**, v. 61, n. 3, p. 340–344, 2014.

CORTES-PENFIELD, N. W.; RAMANI, S.; ESTES, M. K.; ATMAR, R. L. Prospects and Challenges in the Development of a Norovirus Vaccine. **Clinical Therapeutics**, v. 39, n. 8, p. 1537–1549, 2017.

DÁBILLA, N.; NUNES VIEIRA ALMEIDA, T.; CARVALHO REBOUÇAS OLIVEIRA, A.; KIPNIS, A.; NERES SILVA, T.; SOUZA FIACCADORI, F.; TEIXEIRA DE SOUSA, T.; PAULA CARDOSO, D. DAS D. DE; SOUZA, M. Norovirus in feces and nasopharyngeal swab of children with and without acute gastroenteritis symptoms: First report of GI.5 in Brazil and GI.3 in nasopharyngeal swab. **Journal of Clinical Virology**, v. 87, p. 60–66, 2017.

DAI, Y. CHUN; HU, G. FANG; ZHANG, X. FU; SONG, C. LEI; XIANG, W. LONG; WU, X. BO; WANG, L. YI; JIANG, X.; NIE, J. Molecular epidemiology of norovirus gastroenteritis in children in Jiangmen, China, 2005-2007. **Archives of Virology**, v. 156, n. 9, p. 1641–1646, 2011.

DESAI, R.; HEMBREE, C. D.; HANDEL, A.; MATTHEWS, J. E.; DICKEY, B. W.; MCDONALD, S.; HALL, A. J.; PARASHAR, U. D.; LEON, J. S.; LOPMAN, B. Severe outcomes are associated with genogroup 2 genotype 4 norovirus outbreaks: A systematic literature review. **Clinical Infectious Diseases**, v. 55, n. 2, p. 189–193, 2012.

DOLL, M. K.; GAGNEUR, A.; TAPIÉRO, B.; CHAREST, H.; GONZALES, M.; BUCKERIDGE, D. L.; QUACH, C. Temporal Changes in Pediatric Gastroenteritis after Rotavirus Vaccination in Quebec. **Pediatric Infectious Disease Journal**, v. 35, n. 5, p. 555–560, 2016.

EDEN, J. S.; BULL, R. A.; TU, E.; MCIVER, C. J.; LYON, M. J.; MARSHALL, J. A.; SMITH, D. W.; MUSTO, J.; RAWLINSON, W. D.; WHITE, P. A. Norovirus GII.4 variant 2006b caused epidemics of acute gastroenteritis in Australia during 2007 and 2008. **Journal of Clinical Virology**, v. 49, n. 4, p. 265–271, 2010.

ELLIOTT, E. J. Acute gastroenteritis in children. **Bmj**, v. 334, n. 7583, p. 35–40, 2007.

FANKHAUSER, R. L.; NOEL, J. S.; MONROE, S. S.; ANDO, T.; GLASS, R. I. Molecular Epidemiology of “Norwalk- like Viruses” in Outbreaks of Gastroenteritis in the United States. **The Journal of Infectious Diseases**, v. 178, n. 6, p. 1571–1578, 1998.

FIRE CLARK, H.; BORIAN, F. E.; BELL, L. M.; MODESTO, K.; GOUVEA, V.; PLOTKIN, S. A. Protective effect of wc3 vaccine against rotavirus diarrhea in infants during a predominantly serotype 1 rotavirus season. **Journal of Infectious Diseases**, v. 158, n. 3, p. 570–587, 1988.

FREEDMAN, S. B. *et al.* Prospective Assessment of Practice Pattern Variations in the Treatment of Pediatric Gastroenteritis. **Pediatrics**, v. 127, n. 2, p. e287–e295, 2011.

FU, J.-G. *et al.* Molecular Epidemiology of Genogroup II Norovirus Infection Among Hospitalized Children With Acute Gastroenteritis in Suzhou (Jiangsu, China) From 2010 to 2013. **Journal of medical virology**, v. 88, p. 954–960, 2016.

GLASS, R.; PARASHAR, U.; ESTES, M. Norovirus Gastroenteritis. **New Engl J Medicine**, v. 361, n. 18, p. 1776–1785, 2009.

GRAY, J. J. *et al.* European multicenter evaluation of commercial enzyme immunoassays for detecting norovirus antigen in fecal samples. **Clinical and Vaccine Immunology**, v. 14, n. 10, p. 1349–1355, 2007.

GUARINO, A.; ASHKENAZI, S.; GENDREL, D.; VECCHIO, A. LO; SHAMIR, R.; SZAJEWSKA, H. European society for pediatric gastroenterology, hepatology, and nutrition/european society for pediatric infectious diseases evidence-based guidelines for the management of acute gastroenteritis in children in Europe: Update 2014. **Journal of Pediatric Gastroenterology and Nutrition**, v. 59, n. 1, p. 132–152, 2014.

GURGEL, R. G. *et al.* Incidence of Rotavirus and All-Cause Diarrhea in Northeast Brazil Following the Introduction of a National Vaccination Program. **Gastroenterology**, v. 137, n. 6, p. 1970–1975, 2009.

GURGEL, R. Q.; ILOZUE, C.; CORREIA, J. B.; CENTENARI, C.; OLIVEIRA, S. M. T.; CUEVAS, L. E. Impact of rotavirus vaccination on diarrhoea mortality and hospital admissions in Brazil. **Tropical Medicine and International Health**, v. 16, n. 9, p. 1180–1184, 2011.

- HALAIHEL, N.; LIÉVIN, V.; BALL, J. M.; ESTES, M. K.; ALVARADO, F.; VASSEUR, M. Direct inhibitory effect of rotavirus NSP4(114-135) peptide on the Na⁽⁺⁾-D-glucose symporter of rabbit intestinal brush border membrane. **Journal of virology**, v. 74, n. 20, p. 9464–70, 2000.
- HAN, J.; JI, L.; SHEN, Y.; WU, X.; XU, D.; CHEN, L. Emergence and predominance of norovirus GII.17 in Huzhou, China, 2014-2015. **Virology Journal**, v. 12, n. 1, p. 1–7, 2015.
- HARTLING, L.; BELLEMARE, S.; WIEBE, N.; RUSSELL, K. F.; KLASSSEN, T. P.; CRAIG, W. R. Oral versus intravenous rehydration for treating dehydration due to gastroenteritis in children. **Cochrane Database of Systematic Reviews**, n. 1, 2006.
- HEMMING, M.; RÄSÄNEN, S.; HUHTI, L.; PALONIEMI, M.; SALMINEN, M.; VESIKARI, T. Major reduction of rotavirus, but not norovirus, gastroenteritis in children seen in hospital after the introduction of RotaTeq vaccine into the National Immunization Programme in Finland. **European Journal of Pediatrics**, v. 172, n. 6, p. 739–746, 2013.
- HOA-TRAN, T. N.; NAKAGOMI, T.; SANO, D.; SHERCHAND, J. B.; PANDEY, B. D.; CUNLIFFE, N. A.; NAKAGOMI, O. Molecular epidemiology of noroviruses detected in Nepalese children with acute diarrhea between 2005 and 2011: Increase and predominance of minor genotype GII.13. **Infection, Genetics and Evolution**, v. 30, p. 27–36, 2015.
- ITURRIZA-GÓMARA, M.; ELLIOT, A. J.; DOCKERY, C.; FLEMING, D. M.; GRAY, J. J. Structured surveillance of infectious intestinal disease in pre-school children in the community: “The Nappy Study”. **Epidemiology and Infection**, v. 137, n. 7, p. 922–931, 2009.
- JIN, M. *et al.* Emergence of the GII4/2006b Variant and Recombinant Noroviruses in China. **Journal of medical virology**, v. 80, p. 1997–2004, 2008.
- KARAFILLAKIS, E.; HASSOUNAH, S.; ATCHISON, C. Effectiveness and impact of rotavirus vaccines in Europe, 2006-2014. **Vaccine**, v. 33, n. 18, p. 2097–2107, 2015.

KIM, L.; LIEBOWITZ, D.; LIN, K.; KASparek, K.; PASETTI, M. F.; GARG, S. J.; GOTTLIEB, K.; TRAGER, G.; TUCKER, S. N. Safety and immunogenicity of an oral tablet norovirus vaccine, a phase I randomized, placebo-controlled trial. **JCI Insight**, v. 3, n. 13, p. 0–12, 2018.

KITTIGUL, L.; POMBUBPA, K.; TAWEEKATE, Y.; YEEPHOO, T.; KHAMRIN, P.; USHIJIMA, H. Molecular Characterization of Rotaviruses, Noroviruses, Sapovirus, and Adenoviruses in Patients With Acute Gastroenteritis in Thailand. **Journal of medical virology**, v. 81, p. 345–353, 2009.

KOTLOFF, K. L. *et al.* Burden and aetiology of diarrhoeal disease in infants and young children in developing countries (the Global Enteric Multicenter Study, GEMS): A prospective, case-control study. **The Lancet**, v. 382, n. 9888, p. 209–222, 2013.

KRONEMAN, A.; VENNEMA, H.; DEFORCHE, K.; AVOORT, H.; PEÑARANDA, S.; OBERSTE, M. S.; VINJÉ, J.; KOOPMANS, M. An automated genotyping tool for enteroviruses and noroviruses. **Journal of Clinical Virology**, v. 51, n. 2, p. 121–125, 2011.

KULKARNI, R.; PATEL, A.; BHALLA, S.; CHHABRA, P.; CHERIAN, S.; CHITAMBAR, S. D. Characterization of GII.4 noroviruses circulating among children with acute gastroenteritis in Pune, India: 2005-2013. **Infection, Genetics and Evolution**, v. 37, p. 163–173, 2016.

KUMAZAKI, M.; USUKU, S. Norovirus genotype distribution in outbreaks of acute gastroenteritis among children and older people: An 8-year study. **BMC Infectious Diseases**, v. 16, n. 1, p. 1–8, 2016a.

LANATA, C. F.; FISCHER-WALKER, C. L.; OLASCOAGA, A. C.; TORRES, C. X.; ARYEE, M. J.; BLACK, R. E. Global Causes of Diarrheal Disease Mortality in Children <5 Years of Age: A Systematic Review. **PLoS ONE**, v. 8, n. 9, 2013.

LAZZERINI, M.; RONFANI, L. Oral zinc for treating diarrhoea in children (Review) SUMMARY OF FINDINGS FOR THE MAIN COMPARISON. n. 1, 2013.

LEE, I.; KUNTZ, G.; STEVENSON, K. B. Guideline for the Prevention and Control of Norovirus Gastroenteritis. **Prevention**, v. 26, n. 3, p. 1–52, 2013.

LESHEM, E.; MORITZ, R. E.; CURNS, A. T.; ZHOU, F.; TATE, J. E.; LOPMAN, B. A.; PARASHAR, U. D. Rotavirus Vaccines and Health Care Utilization for Diarrhea in the United States (2007-2011). **Pediatrics**, v. 134, n. 1, p. 15–23, 2014.

LIU, L.; OZA, S.; HOGAN, D.; CHU, Y.; PERIN, J.; ZHU, J.; LAWN, J. E.; COUSENS, S.; MATHERS, C.; BLACK, R. E. Global, regional, and national causes of under-5 mortality in 2000–15: an updated systematic analysis with implications for the Sustainable Development Goals. **The Lancet**, v. 388, n. 10063, p. 3027–3035, 2016.

LOGANATHAN, T.; NG, C.-W.; LEE, W.-S.; JIT, M. The Hidden Health and Economic Burden of Rotavirus Gastroenteritis in Malaysia. **The Pediatric Infectious Disease Journal**, v. 35, n. 6, p. 601–606, 2016.

LOPMAN, B. *et al.* Increase in viral gastroenteritis outbreaks in Europe and epidemic spread of new norovirus variant. **Lancet**, v. 363, n. 9410, p. 682–688, 2004.

LOPMAN, B. A.; STEELE, D.; KIRKWOOD, C. D.; PARASHAR, U. D. The Vast and Varied Global Burden of Norovirus: Prospects for Prevention and Control. **PLoS Medicine**, v. 13, n. 4, p. 1–12, 2016.

LU, L.; JIA, R.; ZHONG, H.; XU, M.; SU, L.; CAO, L.; DONG, Z.; DONG, N.; XU, J. Molecular characterization and multiple infections of rotavirus, norovirus, sapovirus, astrovirus and adenovirus in outpatients with sporadic gastroenteritis in Shanghai, China, 2010–2011. **Archives of Virology**, v. 160, n. 5, p. 1229–1238, 2015.

MACGILLIVRAY; FAHEY; MCGUIRE. Lactose avoidance for young children with acute diarrhoea. **Cochrane Database of Systematic Reviews**, n. 1, p. N.PA-N.PA, 2013.

MANS, J.; MURRAY, T. Y.; NADAN, S.; NETSHIKWETA, R.; PAGE, N. A.; TAYLOR, M. B. Norovirus diversity in children with gastroenteritis in South Africa from 2009 to 2013: GII.4 variants and recombinant strains predominate. **Epidemiology and Infection**, v. 144, n. 5, p. 907–916, 2016.

MAWATARI, M.; KATO, Y. Norovirus Gastroenteritis. **Emerging Infectious Diseases: Clinical Case Studies**, p. 203–212, 2014.

MCATEE, C. L. *et al.* Burden of norovirus and rotavirus in children after rotavirus vaccine introduction, Cochabamba, Bolivia. **American Journal of Tropical Medicine and Hygiene**, v. 94, n. 1, p. 212–217, 2016.

MELHEM, N. M. *et al.* Clinical and epidemiological characteristics of norovirus gastroenteritis among hospitalized children in Lebanon. **World Journal of Gastroenterology**, v. 22, n. 48, p. 10557–10565, 2016.

MINISTÉRIO DA SAÚDE. **Acolhimento à demanda espontânea : queixas mais comuns na Atenção Básica**. [s.l: s.n.], v. II

MOYO, S.; HANEVIK, K.; BLOMBERG, B.; KOMMEDAL, O.; VAINIO, K.; MASELLE, S.; LANGELAND, N. Genetic diversity of norovirus in hospitalised diarrhoeic children and asymptomatic controls in Dar es Salaam, Tanzania. **Infection, Genetics and Evolution**, v. 26, p. 340–347, 2014.

MUHSEN, K.; KASSEM, E.; RUBINSTEIN, U.; SCHACHTER, Y.; KREMER, A.; GOREN, S.; ZILBERSTEIN, I.; EPHROS, M.; COHEN, D.; SHULMAN, L. M. Incidence and characteristics of sporadic norovirus gastroenteritis associated with hospitalization of children less than 5 years of age in Israel. **Pediatric Infectious Disease Journal**, v. 32, n. 6, p. 688–690, 2013.

MY, P. V. T. *et al.* Endemic norovirus infections in children, Ho Chi Minh City, Vietnam, 2009–2010. **Emerging infectious diseases**, v. 19, n. 6, p. 29–32, 2013.

NASAB, S. D. M.; SABAHI, F.; MAKVANDI, M.; SAMIEE, S. M.; NADJI, S. A.; RAVANSHAD, M. Epidemiology of rotavirus-norovirus co-infection and determination of norovirus genogrouping among children with acute gastroenteritis in Tehran, Iran. **Iranian Biomedical Journal**, v. 20, n. 5, p. 280–286, 2016.

NATARAJU, S. M. *et al.* Emergence of noroviruses homologous to strains reported from Djibouti (horn of Africa), Brazil, Italy, Japan and USA among children in Kolkata, India. **European Review for Medical and Pharmacological Sciences**, v. 14, n. 9, p. 789–794, 2010.

NAYAK, M. K.; CHATTERJEE, D.; NATARAJU, S. M.; PATIVADA, M.; MITRA, U.; CHATTERJEE, M. K.; SAHA, T. K.; SARKAR, U.; KRISHNAN, T. A new variant of Norovirus GII.4/2007 and inter-genotype recombinant strains of NVGII causing acute watery diarrhoea among children in Kolkata, India. **Journal of Clinical Virology**, v. 45, n. 3, p. 223–229, 2009.

NEVILLE, K. A.; VERGE, C. F.; ROSENBERG, A. R.; O'MEARA, M. W.; WALKER, J. L. Isotonic is better than hypotonic saline for intravenous rehydration of children with gastroenteritis: A prospective randomised study. **Archives of Disease in Childhood**, v. 91, n. 3, p. 226–232, 2006. ORGANIZATION, W. H. The Treatment of Diarrhoea. **A Manual for physicians and other senior health workers**, p. 1–50, 2005.

OROZCO, M.; VASQUEZ, J.; PEDREIRA, C.; OLIVEIRA, L. H. DE; AMADOR, J. J.; MALESPIN, O.; ANDRUS, J.; TATE, J.; PARASHAR, U.; PATEL, M. Uptake of Rotavirus Vaccine and National Trends of Acute Gastroenteritis among Children in Nicaragua. **The Journal of Infectious Diseases**, v. 200, n. s1, p. S125–S130, 2009.

PAYNE, D. C. *et al.* Norovirus and Medically Attended Gastroenteritis in U.S. Children. **New England Journal of Medicine**, v. 368, n. 12, p. 1121–1130, 2013a.

PEDIATRIA, S. B. DE. **Tratado de pediatria de Nelson.** Third ed. Barueri: Manole, 2014.

PUUSTINEN, L.; BLAZEVIC, V.; HUHTI, L.; SZAKAL, E. D.; HALKOSALO, A.; SALMINEN, M.; VESIKARI, T. Norovirus genotypes in endemic acute gastroenteritis of infants and children in Finland between 1994 and 2007. **Epidemiology and Infection**, v. 140, n. 2, p. 268–275, 2012.

QAZOUI, M. EL; OUMZIL, H.; BAASSI, L.; OMARI, N. EL; SADKI, K.; AMZAZI, S.; BENHAFID, M.; AOUAD, R. EL. Rotavirus and norovirus infections among acute gastroenteritis children in Morocco. **BMC Infectious Diseases**, v. 14, n. 1, p. 1–9, 2014.

QIAO, N.; WANG, X. Y.; LIU, L. Temporal evolutionary dynamics of norovirus GII.4 variants in China between 2004 and 2015. **PLoS ONE**, v. 11, n. 9, p. 1–16, 2016.

RABONI, S. M.; DAMASIO, G. A. C.; FERREIRA, C. E.; PEREIRA, L. A.; NOGUEIRA, M. B.; VIDAL, L. R.; CRUZ, C. R.; ALMEIDA, S. M. Acute gastroenteritis and enteric viruses in hospitalised children in southern Brazil: aetiology, seasonality and clinical outcomes. **Memórias do Instituto Oswaldo Cruz**, v. 109, n. 4, p. 428–435, jul. 2014.

RAHMAN, M.; RAHMAN, R.; NAHAR, S.; HOSSAIN, S.; AHMED, S.; FARUQUE, A. S. G.; AZIM, T. Norovirus Diarrhea in Bangladesh, 2010–2014: Prevalence, Clinical Features, and Genotypes. **Journal of medical virology**, v. 88, p. 1742–1750, 2016.

RAMIREZ, S.; GIAMMANCO, G.; GRAZIA, S. DE; COLOMBA, C.; MARTELLA, V.; ARISTA, S. Emerging GII.4 Norovirus Variants Affect Children With Diarrhea in Palermo, Italy in 2006. **Journal of medical virology**, v. 81, p. 139–145, 2009.

RHEINGANS, R. D.; ANTIL, L.; DREIBELBIS, R.; PODEWILS, L. J.; BRESEE, J. S.; PARASHAR, U. D. Economic Costs of Rotavirus Gastroenteritis and Cost-Effectiveness of Vaccination in Developing Countries. **The Journal of Infectious Diseases**, v. 200, n. s1, p. S16–S27, 2009.

RICHARDSON, V.; HERNANDEZ-PICHARDO, J.; QUINTANAR-SOLARES, M.; ESPARZA-AGUILAR, M.; JOHNSON, B.; GOMEZ-ALTAMIRANO, C. M.; PARASHAR, U.; PATEL, M. Effect of Rotavirus Vaccination on Death from Childhood Diarrhea in Mexico. **New England Journal of Medicine**, v. 362, n. 4, p. 299–305, 2010.

RIERA-MONTES, M.; O'RYAN, M.; VERSTRAETEN, T. Norovirus and Rotavirus disease severity in children. **The Pediatric Infectious Disease Journal**, v. 37, n. 6, p. 1, 2017.

RODRIGUES, L. SBP: Diarreia aguda - diagnóstico e tratamento. **Gastroenterologia, Departamento Científico De Silva**, p. 1–15, 2017.

RUUSKA, T.; VESIKARI, T. Rotavirus disease in Finnish children: use of numerical scores for clinical severity of diarrhoeal episodes. **Scandinavian journal of infectious diseases**, v. 22, n. 3, p. 259–67, 1990.

SÁFADI, M. A. P.; BEREZIN, E. N.; MUNFORD, V.; ALMEIDA, F. J.; MORAES, J. C. DE; PINHEIRO, C. F.; RACZ, M. L. Hospital-based surveillance to evaluate the impact of rotavirus vaccination in São Paulo, Brazil. **Pediatric Infectious Disease Journal**, v. 29, n. 11, p. 1019–1022, 2010.

SANTOS, V. S. *et al.* Acute norovirus gastroenteritis in children in a highly rotavirus-vaccinated population in Northeast Brazil. **Journal of Clinical Virology**, v. 88, p. 33–38, 2017.

SCHMID, M.; OEHME, R.; SCHALASTA, G.; BROCKMANN, S.; KIMMIG, P.; ENDERS, G. Fast detection of Noroviruses using a real-time PCR assay and automated sample preparation. **BMC.Infect.Dis.**, v. 4, p. 15, 2004.

SHARIA M AHMED, ARON J HALL, ANNE E ROBINSON, LINDA VERHOEF, PRASANNA PREMKUMAR, UMESH D PARASHAR, MARION KOOPMANS, B. A. L. Global prevalence of norovirus in cases of gastroenteritis: a systematic review and meta-analysis. **The Lancet**

Infectious Diseases, v. 14, p. 725–730, 2014.

SHE, R. C.; HYMAS, W. C.; TAGGART, E. W.; PETTI, C. A.; HILLYARD, D. R. Performance of enterovirus genotyping targeting the VP1 and VP2 regions on non-typeable isolates and patient specimens. **Journal of Virological Methods**, v. 165, n. 1, p. 46–50, 2010.

SIEBENGA, J. J. *et al.* Norovirus Illness Is a Global Problem: Emergence and Spread of Norovirus GII.4 Variants, 2001–2007. **The Journal of Infectious Diseases**, v. 200, n. 5, p. 802–812, 2009.

SIEBENGA, J. J.; VENNEMA, H.; RENCKENS, B.; BRUIN, E. DE; VEER, B. VAN DER; SIEZEN, R. J.; KOOPMANS, M. Epochal Evolution of GGII.4 Norovirus Capsid Proteins from 1995 to 2006. **Journal of Virology**, v. 81, n. 18, p. 9932–9941, 2007.

STRAND, T. A.; SHARMA, P. R.; GJESSING, H. K.; ULAK, M.; CHANDYO, R. K.; ADHIKARI, R. K.; SOMMERFELT, H. Risk factors for extended duration of acute diarrhea in young children. **PLoS ONE**, v. 7, n. 5, p. 3–8, 2012.

TAN, D.; DENG, L.; WANG, M.; LI, X.; MA, Y.; LIU, W. High Prevalence and Genetic Diversity of Noroviruses Among Children With Sporadic Acute Gastroenteritis in Nanning City, China, 2010–2011. **Journal of medical virology**, v. 87, p. 498–503, 2015.

THONGPRACHUM, A.; KHAMRIN, P.; CHAN-IT, W.; MALASAO, R.; CHAIMONGKOL, N.; OKITSU, S.; MIZUGUCHI, M.; MANEEKARN, N.; HAYAKAWA, S.; USHIJIMA, H. Emergence of Norovirus GII/4 2006a and 2006b variants in hospitalized children with acute gastroenteritis in Thailand. **Clinical Laboratory**, v. 59, n. 3–4, p. 271–276, 2013.

TRANG, N. V; CHOISY, M.; NAKAGOMI, T.; CHINH, N. T. M.; DOAN, Y. H.; YAMASHIRO, T.; BRYANT, J. E.; NAKAGOMI, O.; ANH, D. D. Determination of cut-off cycle threshold values in routine RT-PCR assays to assist differential diagnosis of norovirus in children hospitalized for acute gastroenteritis. **Epidemiology and infection**, v. 143, n. 15, p. 3292–9, 2015.

TRANG, N. V; LUAN, L. T.; KIM-ANH, L. T.; HAU, V. T. B.; NHUNG, L. T. H. Detection and Molecular Characterization of Noroviruses and Sapoviruses in Children Admitted to Hospital With Acute Gastroenteritis in Vietnam. **Journal of medical virology**, v. 84, p. 290–297, 2012.

TROEGER, H.; LODDENKEMPER, C.; SCHNEIDER, T.; SCHREIER, E.; EPPEL, H.-J.; ZEITZ, M.; FROMM, M.; SCHULZKE, J.-D. Structural and functional changes of the duodenum in human norovirus infection. **Gut**, v. 58, n. 8, p. 1070–1077, 2008.

UNITED NATIONS. **United Nations Millennium Declaration**, 2000.

VANDENPLAS, Y. *et al.* Cost/benefit of synbiotics in acute infectious gastroenteritis: Spend to save. **Beneficial Microbes**, v. 3, n. 3, p. 189–194, 2012.

VECCHIO, A. LO; ZACUR, G. M. Clostridium difficile infection: An update on epidemiology, risk factors, and therapeutic options. **Current Opinion in Gastroenterology**, v. 28, n. 1, p. 1–9, 2012.

VICTORA, C. G.; BARROS, A. J. D. Effect of breastfeeding on infant and child mortality due to infectious diseases in less developed countries: A pooled analysis. **Lancet**, v. 355, n. 9202, p. 451–455, 2000.

WHO. Rotavirus vaccines:an update. **Weekly epidemiological record / Health Section of the Secretariat of the League of Nations**, v. 84, n. 50, p. 533–540, 2009.

YANG, S. Y.; HWANG, K. P.; WU, F. T.; WU, H. S.; HSIUNG, C. A.; CHANG, W. C.; LIN, J. S.; YANG, S. C.; HUANG, S. L.; HUANG, Y. C. Epidemiology and Clinical Peculiarities of Norovirus and Rotavirus Infection in Hospitalized Young Children with Acute Diarrhea in Taiwan, 2009. **Journal of Microbiology, Immunology and Infection**, v. 43, n. 6, p. 506–514, 2010.

YEN, C.; WIJKSWO, M. E.; LOPMAN, B. A.; VINJE, J.; PARASHAR, U. D.; HALL, A. J. Impact of an Emergent Norovirus Variant in 2009 on Norovirus Outbreak Activity in the United States. **Clinical Infectious Diseases**, v. 53, n. 6, p. 568–571, 2011.

ZHANG, P.; CHEN, L.; FU, Y.; JI, L.; WU, X.; XU, D.; HAN, J. Clinical and molecular analyses of norovirus-associated sporadic acute gastroenteritis: The emergence of GII.17 over GII.4, Huzhou, China, 2015. **BMC Infectious Diseases**, v. 16, n. 1, p. 1–8, 2016.

ZHANG, S. *et al.* Symptomatic and Asymptomatic Infections of Rotavirus, Norovirus, and Adenovirus Among Hospitalized Children in Xi'an, China. **Journal of Medical Virology**, v. 83, p. 1476–1484, 2011.

ZHANG, S. X.; LI, L.; YIN, J. W.; JIN, M.; KONG, X. Y.; PANG, L. L.; ZHOU, Y. K.; TIAN, L. G.; CHEN, J. X.; ZHOU, X. N. Emergence of human caliciviruses among diarrhea cases in southwest China. **BMC Infectious Diseases**, v. 16, n. 1, p. 1–9, 2016.

ZHENG, D. P.; ANDO, T.; FANKHAUSER, R. L.; BEARD, R. S.; GLASS, R. I.; MONROE, S. S. Norovirus classification and proposed strain nomenclature. **Virology**, v. 346, n. 2, p. 312–323, 2006.

ZHIRAKOVSKAIA, E.; TIKUNOV, A. Y.; BODNEV, S. A.; KLEMESHEVA, V. V.; NETESOV, S. V.; TIKUNOVA, N. V. Molecular Epidemiology of Noroviruses Associated With Sporadic Gastroenteritis in Children in Novosibirsk, Russia, 2003–2012. **Journal of medical virology**, v. 87, p. 740–753, 2015.

3. NORMAS PARA PUBLICAÇÃO

3.1. JOURNAL INFORMATION

The world's first multidisciplinary Open Access journal, *PLOS ONE* accepts scientifically rigorous research, regardless of novelty. *PLOS ONE*'s broad scope provides a platform to publish primary research, including interdisciplinary and replication studies as well as negative results. The journal's publication criteria are based on high ethical standards and the rigor of the methodology and conclusions reported.

3.2. SCOPE

PLOS ONE features reports of original research from the natural sciences, medical research, engineering, as well as the related social sciences and humanities that will contribute to the base of scientific knowledge. By not excluding research on the basis of subject area, *PLOS ONE* facilitates the discovery of connections between research whether within or between disciplines.

3.3. SYSTEMATIC REVIEWS AND META-ANALYSES

A systematic review paper, as defined by The Cochrane Collaboration, is a review of a clearly formulated question that uses explicit, systematic methods to identify, select, and critically appraise relevant research, and to collect and analyze data from the studies that are included in the review. These reviews differ substantially from narrative-based reviews or synthesis articles. Statistical methods (meta-analysis) may or may not be used to analyze and summarize the results of the included studies.

Reports of systematic reviews and meta-analyses must include a completed PRISMA (Preferred Reporting Items for Systematic Reviews and Meta-Analyses) checklist and flow diagram to accompany the main text.

Authors must also state in their “Methods” section whether a protocol exists for their systematic review, and if so, provide a copy of the protocol as supporting information and provide the registry number in the abstract.

If your article is a systematic review or a meta-analysis you should:

- State this in your cover letter
- Select “Research Article” as your article type when submitting
- Include the PRISMA flow diagram as Fig 1 (required where applicable)
- Include the PRISMA checklist as supporting information

3.4. CRITERIA FOR PUBLICATION

A. The study presents the results of primary scientific research

PLOS ONE is designed to communicate primary scientific research. We welcome submissions in the natural sciences, medical research, engineering, as well as the related social sciences and humanities that will contribute to the base of scientific knowledge.

We will not consider:

- Reviews
- Case reports
- Study protocols
- Hypothesis or proposal papers
- Letters, commentaries, or essays
- Opinion pieces
- Policy papers
- Clinical practice guidelines
- Any other type of secondary literature
- Monographs

We will occasionally commission Collection Reviews or Overviews, but these articles are associated with specific, pre-planned Collections and will not be considered unless solicited.

We will consider the following article types:

<i>Systematic reviews</i>	We consider publishing systematic reviews only if the methods ensure the comprehensive and unbiased sampling of existing literature.
<i>Submissions describing methods, software, databases, or other tools</i>	We consider submissions describing methods, software, databases, or other tools if they follow the appropriate reporting guidelines.
<i>Qualitative research</i>	We consider publishing qualitative research only if it adheres to appropriate study design and reporting guidelines.
<i>Studies reporting negative results</i>	

B. Results reported have not been published elsewhere

Previously Published Studies

PLOS ONE does not accept for publication studies that have already been published, in whole or in part, elsewhere in the peer-reviewed literature. All figures included in manuscripts should be original, and should not have been published in any previous publications.

In addition, we will not consider submissions that are currently under consideration for publication elsewhere.

PLOS supports authors who wish to share their work early through deposition of manuscripts in preprint servers. This does not impact consideration of the manuscript at any PLOS journal. We will consider manuscripts that have been deposited in preprint servers such as bioRxiv or arXiv, published as a thesis, or presented at conferences.

Replication Studies

If a submitted study replicates or is very similar to previous work, authors must provide a sound scientific rationale for the submitted work and clearly reference and discuss the existing literature. Submissions that replicate or are derivative of existing work will likely be rejected if authors do not provide adequate justification.

C. Experiments, statistics, and other analyses are performed to a high technical standard and are described in sufficient detail.

Experiments must have been conducted rigorously, with appropriate controls and replication. Sample sizes must be large enough to produce robust results, where applicable. Methods and reagents must be described in sufficient detail for another researcher to reproduce the experiments described.

D. Conclusions are presented in an appropriate fashion and are supported by the data

The data presented in the manuscript must support the conclusions drawn. Submissions will be rejected if the interpretation of results is unjustified or inappropriate, so authors should avoid overstating their conclusions. Authors may discuss possible implications for their results as long as these are clearly identified as hypotheses instead of conclusions.

E. The article is presented in an intelligible fashion and is written in standard English

PLOS ONE does not copyedit accepted manuscripts, so the language in submitted articles must be clear, correct, and unambiguous. We may reject papers that do not meet these standards.

If the language of a paper is difficult to understand or includes many errors, we may recommend that authors seek independent editorial help before submitting a revision.

F. The research meets all applicable standards for the ethics of experimentation and research integrity

Ethics of Experimentation

Research published in *PLOS ONE* must have been conducted to the highest ethical standards. We reserve the right to reject any submission that does not meet these standards, which in some cases are more stringent than local ethical standards.

Approval from the relevant body is required for studies involving:

- Humans (live or tissue), including studies that are observational, survey-based, or include any personal data.
- Animals (live or tissue), including observational studies
- Cell lines that are not commercially available
- Field sampling
- Potential biosafety implications

If approval was not obtained, authors must explain why it was not required.

Publication Ethics

PLOS ONE is a member of the Committee on Publication Ethics (COPE). We abide by its Code of Conduct and aim to adhere to its Best Practice Guidelines. Authors are expected to comply with best practices in publication ethics, specifically regarding authorship, dual publication, plagiarism, figure manipulation, and competing interests.

G. The article adheres to appropriate reporting guidelines and community standards for data availability.

Reporting Guidelines

Results must be rigorously reported, as appropriate based on community standards.

Data Availability

Authors must follow standards and practice for data deposition in publicly available resources including those created for gene sequences, microarray expression, structural studies, and similar kinds of data. Failure to comply with community standards may result in rejection.

3.5. STYLE AND FORMAT

File format	Manuscript files can be in the following formats: DOC, DOCX, or RTF. Microsoft Word documents should not be locked or protected.
Length	LaTeX manuscripts must be submitted as PDFs.
Font	Manuscripts can be any length. There are no restrictions on word count, number of figures, or amount of supporting information.
Headings	We encourage you to present and discuss your findings concisely.
Layout and spacing	Use a standard font size and any standard font, except for the font named “Symbol”. To add symbols to the manuscript, use the Insert → Symbol function in your word processor or paste in the appropriate Unicode character.
Page and line numbers	Limit manuscript sections and sub-sections to 3 heading levels. Make sure heading levels are clearly indicated in the manuscript text.
Footnotes	Manuscript text should be double-spaced.
Language	Do not format text in multiple columns.
Abbreviations	Include page numbers and line numbers in the manuscript file. Use continuous line numbers (do not restart the numbering on each page).
Reference style	Footnotes are not permitted. If your manuscript contains footnotes, move the information into the main text or the reference list, depending on the content.
Equations	Manuscripts must be submitted in English.
	You may submit translations of the manuscript or abstract as supporting information.
	Define abbreviations upon first appearance in the text.
	Do not use non-standard abbreviations unless they appear at least three times in the text.
	Keep abbreviations to a minimum.
	PLOS uses “Vancouver” style
	We recommend using MathType for display and inline equations, as it will provide the most reliable outcome. If this

is not possible, Equation Editor or Microsoft's Insert→Equation function is acceptable.

Avoid using MathType, Equation Editor, or the Insert→Equation function to insert single variables (e.g., “ $a^2 + b^2 = c^2$ ”), Greek or other symbols (e.g., β , Δ , or ' [prime]), or mathematical operators (e.g., x , \geq , or \pm) in running text. Wherever possible, insert single symbols as normal text with the correct Unicode (hex) values.

Do not use MathType, Equation Editor, or the Insert→Equation function for only a portion of an equation. Rather, ensure that the entire equation is included. Equations should not contain a mix of different equation tools. Avoid “hybrid” inline or display equations, in which part is text and part is MathType, or part is MathType and part is Equation Editor.

Nomenclature	Use correct and established nomenclature wherever possible.
<i>Units of measurement</i>	Use SI units. If you do not use these exclusively, provide the SI value in parentheses after each value.
<i>Drugs</i>	Provide the Recommended International Non-Proprietary Name (rINN).
<i>Species names</i>	Write in italics (e.g., <i>Homo sapiens</i>). Write out in full the genus and species, both in the title of the manuscript and at the first mention of an organism in a paper. After first mention, the first letter of the genus name followed by the full species name may be used (e.g., <i>H. sapiens</i>).
<i>Genes, mutations, genotypes, and alleles</i>	Write in italics. Use the recommended name by consulting the appropriate genetic nomenclature database (e.g., HUGO for human genes). It is sometimes advisable to indicate the synonyms for the gene the first time it appears in the text. Gene prefixes such as those used for oncogenes or cellular

<i>Allergens</i>	localization should be shown in roman typeface (e.g., v-fes, c-MYC).
	The systematic allergen nomenclature of the World Health Organization/International Union of Immunological Societies (WHO/IUIS) Allergen Nomenclature Subcommittee should be used for manuscripts that include the description or use of allergenic proteins. For manuscripts describing new allergens, the systematic name of the allergen should be approved by the WHO/IUIS Allergen Nomenclature Subcommittee prior to manuscript publication..

Manuscript Organization

Beginning section	<i>The following elements are required, in order:</i>
	<ul style="list-style-type: none"> • Title page: List title, authors, and affiliations as first page of manuscript • Abstract • Introduction
Middle section	<i>The following elements can be renamed as needed and presented in any order:</i>
	<ul style="list-style-type: none"> • Materials and Methods • Results • Discussion • Conclusions (optional)
Ending section	<i>The following elements are required, in order:</i>
	<ul style="list-style-type: none"> • Acknowledgments • References

	<ul style="list-style-type: none">• Supporting information captions (if applicable)
Other elements	<ul style="list-style-type: none">• Figure captions are inserted immediately after the first paragraph in which the figure is cited. Figure files are uploaded separately.• Tables are inserted immediately after the first paragraph in which they are cited.• Supporting information files are uploaded separately.

4. ARTIGO ORIGINAL**GLOBAL CIRCULATION OF NOROVIRUS GENOTYPES IN YOUNG CHILDREN
OVER TEN YEARS: SYSTEMATIC REVIEW AND META-ANALYSIS.**

Hiram Menezes N. Filho¹, Carlos Henrique F. Oliveira¹, Paulo R. S. Martins-Filho², Victor S. Santos³, Ricardo Q. Gurgel^{1*}

¹Department of Medicine. Federal University of Sergipe, Aracaju, Brazil.

²Investigative Pathology Laboratory. Federal University of Sergipe, Aracaju, Brazil.

³Centre for Epidemiology and Public Health. Federal University of Alagoas, Arapiraca, Brazil.

*Ricardo Q. Gurgel.

Email: [ricardoqgurgel@gmail.com \(RG\)](mailto:ricardoqgurgel@gmail.com (RG))

Abstract

Introduction: Norovirus has emerged as the leading etiologic agent in acute gastroenteritis in young children since the introduction of vaccines against Rotavirus. Norovirus most predominant genotypes change every 2-3 years.

Methods: We describe the temporal dynamics involving Norovirus genotypes over ten years through a systematic review and meta-analysis of studies published between 2006 and 2015. 108 studies were included to evaluate the burden of Norovirus-associated acute gastroenteritis in children younger than 5 years of age. 51 studies provided information to estimate the proportion of Norovirus circulating genotypes into three periods 2006-2008, 2009-2011 and 2012-2015 using meta-analysis.

Results: Over the period of 10 years, 7,072 specimens of Norovirus have been genotyped from stool samples of children less than 5 years of age. Genogroup II responded to most of specimens over the three periods in America, Africa, Asia and Europe, ranging from 89.2% (95% CI, 81.8–93.4) in America 2006-2008 to 98.3% (95% CI, 96.1–99.3) in Asia 2009-2011. Genotype GII.4 presented as the most predominant genotype, accounting for the lowest proportion in Africa 2009-2011 41% (95% CI, 35.1-46.2) and the highest in America 2012-2015 80% (95% CI, 67.5-89.0).

Discussion: NoV GII.4 is the most common genotype involved in pathogenesis of NoV-associated acute gastroenteritis in children <5. Genotypes GII.3, GII.6 and GII.12 also respond to a great amount of NoV infections varying accord periods and continents.

Keywords: Epidemiology; Acute gastroenteritis; Norovirus; Genotypes.

Introduction

Despite substantial efforts in prevention and management of acute gastroenteritis (AGE), diarrheal disease remains as a leading cause of death in children less than 5 years old. Even though AGE is preventable and treatable, annually it accounts for 526.00 deaths in this age group [1].

Since the introduction of Rotavirus (RoV) vaccines in national immunization programs in developed and developing countries, overall children mortality and hospitalizations due to diarrhea as well as RoV-associated AGE have declined [2–5]. On the other hand, Norovirus (NoV), that used to be the second leading etiologic agent of AGE in children <5, has assumed a pivotal role in this pathology [6–11]. According to a recent global meta-analysis comprising studies published between January 2008 and March 2014, NoV responds to 18% of AGE cases [12]. NoV- associated AGE usually occurs in outbreaks during the year round, affecting all age groups, especially young children and the elderly. Due to fecal-oral mode of transmission, low infectious doses and extended shedding, NoV reinforces itself as a common cause of AGE [13]. Besides, NoV is genetically classified into 6 genogroups, composed by several genotypes, 9GI, 22 GII and 2 GIV according to capsid gene. The genotype GII.4 is responsible for the majority of NoV cases featuring some fashion that every 2-3 years a new GII.4 variant emerges and it becomes the predominant strain [14].

Although RoV infection mortality is higher, NoV also causes severe to moderate impact on morbidity, direct health costs and productivity losses. It is estimated that AGE in children < 5 costs around \$39.8 billion [15–17]. Once this age group is the most vulnerable for severity, it deserves particular interest in prevention. Experimental vaccines against NoV have not shown satisfactory results [18], therefore most of the disease control strategy focus on hand hygiene, environmental cleaning, personal protective equipment and cases notification [19].

To our knowledge, there are no systematic reviews published in the literature that comprise NoV genotypes circulation in children < 5. Epidemiological characterization of NoV infections is an important aspect for decision-makers when elaborating control strategies against this pathogen (i.e. vaccine development). To understand the pattern of NoV circulation, we conducted a systematic review and meta-analysis to describe NoV circulating genotypes in this age group over ten years.

Methods

Search strategy and selection criteria

We performed a systematic review on MEDLINE (Medical Literature Analysis and Retrieval System Online), SCOPUS and LILACS (Latin American and Caribbean Health Sciences Literature) databases searching for studies published through the period of January 1st 2006 and December 31st 2016 using the following search terms: “Norovirus”, “Norwalk-like virus”, “Caliciviridae infections”; and related terms.

Two independent reviewers (H.N & C.O.) screened titles and abstracts for relevance and selected original articles that included children less than 5 years old with symptoms of AGE, reported data on norovirus genotypes or reported epidemiological information. A third independent reviewer (R.G) evaluated the result of this screening section, selecting the titles for full-text assessment. To ensure the capture of all relevant studies we cross-referenced all articles from the bibliography of the selected articles. After full-text assessment, we selected the articles that met the following criteria: (1) Conducted between 2006 and 2015; (2) English, Spanish or Portuguese as the publication language; (3) Conducted for at least 12 months; (4) Specified the location where the study was performed; (5) Included children less than 5 years old with signs and symptoms of AGE; (6) PCR-based technique for Norovirus infection diagnose from stool samples. We excluded

review or opinion publications without original data; studies that did not provide a denominator (i.e., the total number of patients with AGE in the study population); studies that did not allow the extraction of data for children < 5.

Data extraction and bias assessment

Relevant data was extracted from each article: First author, title, journal, year of publication, start date of samples gathering, end date of samples gathering, country, total number of children less than 5 years old, positive noroviruses cases in children less than 5 years old and noroviruses genotypes isolated from stool samples of infected children. When there was population overlap among two different publications, the one with less time of data collection was excluded.

Two spreadsheets were edited. The first held data of articles that was possible extracting Norovirus AGE incidence in children less than 5 years old. The second held data of articles that was possible extracting Norovirus genotypes isolated from stool samples of children less than 5 years old diagnosed with Norovirus AGE. Moreover, the second spreadsheet comprised all NoV capsid genotypes reported in each study (Genogroups I and II). Each spreadsheet had the studies information divided into three groups according to the period of samples collection (2006-2008, 2009-2011, 2012-2015)

Data synthesis

Proportion of NoV circulating genotypes were calculated using logit transformation. We used the 95% CIs to graphically represents NoV isolated genotypes in children aged 0-5 years according to the geographic region. Statistical heterogeneity was assessed by using the Cochran Q test [20] and

quantified by the I^2 index [21]. Analyzes were performed using R statistical programming language version 2.10.13.

Results

Database search was performed on March 17th, 2017. 6412 records were identified, 3576 from Scopus, 2784 Medline, and 52 LILACS. Afterwards 2011 duplicates were removed, and 4401 titles and abstracts were screened for relevance. 696 full-text articles have been assessed for eligibility, and ultimately 108 articles have been included in systematic review. 51 articles reported NoV genotypes in infected children <5 over the periods of 2006-2008, 2009-2011 and 2012-2015 (Figure 1 and S1 Table). The selected records comprised studies from four continents, including 5 countries in Africa, 4 in America, 16 in Asia, 9 in Europe. It was obtained information on 7,072 NoV specimens from children aged < 5 presenting AGE symptoms over 10 years. NoV most predominant genotypes varied over continents and periods. There were no eligible reports from Oceania identified during the entire period of study.

Fig 1. Flow diagram of study selection.

Table 1. Norovirus specimens by genogroups from 2006 to 2015.

Norovirus circulating genotypes

2006-2008

Twenty-eight studies [(6,22–44)] provided data on NoV circulating genotypes from 2006 to 2008. Most of them were carried out in Asia (20), where 1402 specimens of NoV were genotyped. Europe

had 243 specimens of NoV genotyped, followed by America and Africa reaching 106 and 72 specimens respectively. Proportion of NoV Genogroup I specimens varied from 1.8% (95% CI, 0.5–6.8) in Europe to 10.8% (95% CI, 6.1–18.2) in America, the highest proportion found for this genogroup. Genogroup II specimens varied from 89.2% (95% CI, 81.8–93.4) in Asia to 98.2% (95% CI, 93.2–99.5) in Europe. Genotype GII.4 was the predominant one in the four continents over this period. Its circulation proportion varied from 63% (95% CI, 56.7–68.8) in Europe to 75% (95% CI, 66.5–82.7) in America. Genotype GII.3 presented significant circulation in Asia (17%; 95% CI, 15.3–19.2) and Europe (27%; 95% CI, 22.0–33.1), while accounted for 3% (95% CI, 0.1–8.0) of genotyped specimens in America. Genotype GII.6 also had a significant circulation in Africa (8%; 95% CI, 3.9–17.0) and Europe (4%; 95% CI, 2.3–7.4). Figure 2

Fig 2. Norovirus circulating genotypes from 2006 to 2008.

2009-2011

Twenty-eight studies [\(6–9,26,27,29,31–35,38,41,45–57\)](#) provided data on NoV circulating genotypes from 2009 to 2011. Most of the studies were carried out in Asia (18), where 2027 specimens of NoV were genotyped. America accounted for 463 specimens, while Africa and Europe accounted for 301 and 108 specimens respectively. Genogroup I specimens varied from 1.7% (95% CI, 0.7–3.9) in Asia to 5.1% (95% CI, 2.0–12.3) in Europe. On the other hand, Genogroup II reached the highest rates over the period varying from 94.9% (95% CI, 87.7–98.0) in Europe to 98.3% (95% CI, 96.1–99.3) in Asia. Genotype GII.4 remained as the predominant genotype in the four continents, though just in America it reached circulation proportion higher than 60% (63%; 95% CI, 58.6–67.3). Africa, Asia and Europe registered the widest variety of circulating genotypes over this period. Even though, most of these genotypes were not presented,

GII.3 remained with significant proportion of circulating specimens in Asia (23%; 95% CI, 21.1-24.8), while GII.6 was the third most common circulating genotype in this continent (2.8%; 95% CI, 2.1-3.6). Furthermore, GII.12 emerged as an important genotype in America (8%; 95% CI 5.9-10.8) and Europe (6%; 95% CI, 2.6-11.6).

Fig 3. Norovirus circulating genotypes from 2009 to 2011.

2012-2015

Over the period from 2012 to 2015, only data from America, Africa and Asia has been available from 16 studies [6,31-33,35,38,46,52,53,58-64]. Asia had 2122 specimens genotyped, followed by Africa (177) and America (51). Genogroup I proportion varied from 2.8% (95% CI, 1.2-6.0) in Asia to 7.3% (95% CI, 4.0-12.2) in Africa as Genogroup II varied from 92.7% (95% CI, 87.8-96.0) to 97.3% (95% CI, 94.0-98.8) in Asia. Genotype GII.4 remained as the predominant circulating genotype in the three continents. In America GII.4 accounted for 80% (95% CI, 67.5-89.0) of circulating genotypes, while it accounted for 57% (95% CI, 49.7-64.1) in Africa and 56% (95% CI, 53.5-57.8) in Asia. The Asian and African continent presented the same pattern of most common circulating genotypes. Genotype GII.3 re-emerged its importance once it presented as the second most common in Asia (33%; 95% CI, 30.4-34.3) and Africa (17%; 95% CI, 12.1-23.1). Besides, genotype GII.12 presented as the third most common genotype in Asia (5%; 95% CI, 3.8-5.6) and Africa (6%; 95% CI, 3.1-10.0).

Fig 4. Norovirus circulating genotypes from 2012 to 2015.

Discussion

Norovirus-associated acute gastroenteritis has fomented health public concernment since the decrease of Rotavirus-associated diarrhea. Once NoV has been recognized as the predominant etiologic agent of AGE in children <5, several studies have been carried out to determine clinical and epidemiological patterns resulted from this etiological change (65). To our knowledge this is the first systematic review that globally evaluates NoV genotypes circulation for the age group of children below 5 years of age. This used to be the most affected group by Rotavirus infection, and now NoV is replacing RV as the most frequent diarrhea etiologic agent in most part of the world [6–10].

It is known that NoV-associated outbreaks are influenced by the leading NoV genotype in circulation. This meta-analysis evaluated 7,072 specimens of NoV isolated from stool samples of children from 4 continents. Genogroup II Norovirus responded for the majority of specimens, reaching the lowest proportion in America 2006-2008 (89.2%) and the highest in Asia 2009-2011 (98.3%). Genotype GII.4 responded to more than 50% of circulating genotypes, excepting for 2009-2011 period in Asia, Africa and Europe. This data is consistent with previous review that suggests that hospitalizations and deaths are more likely associated with GII.4 genotype (66).

It is well observed that NoV circulating genotypes vary over time, specially through a 2-3 year period, when another genotype or a novel variant genotype emerges and displaces the dominant virus (67). This meta-analysis identified NoV GII.4 as the most common genotype involved in pathogenesis of NoV-associated AGE. During the period from 2006 to 2008, GII.4 accounted for more than 60% of NoV specimens in Asia, Africa, America and Europe, followed by GII.3, that also played a pivotal role in Asia and Europe. This predominance pattern was threatened by the increased circulation of other genotypes in the period from 2009-2011. During this period GII.4

proportion decreased, mainly in Asia and Europe, where GII.12 arose as an important genotype replacing GII.3 position in the previous period. Besides GII.13, that has been significantly recognized in Asia 2006-2008, emerged as an important genotype in America. During the last period 2012-2015, the pattern observed from 2006 to 2008 returned. GII.4 responded to the most of genotypes, followed by GII.3. Highlights for GII.12, that remained as a dominant genotype in Asia and Africa, and GII.6 that assumed an important circulation in America once it also circulated in Africa and Europe from 2006 to 2008 and Asia from 2009 to 2011.

Information provided by this systematic review.

This is a descriptive study, therefore there is no causal insight that could explain molecular epidemiological changes. Neither all continents have available data² nor all countries are comprised in the three different periods. Besides, not all studies provided NoV genotypes clustering, which limits molecular epidemiological comparison and genotypes variants evolution over the years.

Our study certainly contributes to synthesize epidemiological information regarding NoV genotypes circulation in children < 5. As described, each continent has particular features on the most identified genotypes over the periods. Excepting for GII.4, other genotypes arise or decline as pivotal etiologic specimens. These data enhance the considerations for a vaccine development once it signals that other genotypes should be targeted and that the same vaccine may not be useful for different continents. Finally, additional studies are necessary to increase the knowledge on Noroviruses epidemiological behavior.

References

1. Liu L, Oza S, Hogan D, Chu Y, Perin J, Zhu J, et al. Global, regional, and national causes of under-5 mortality in 2000–15: an updated systematic analysis with implications for the

- Sustainable Development Goals. *Lancet.* 2016;388(10063):3027–35.
2. Doll MK, Gagneur A, Tapiéro B, Charest H, Gonzales M, Buckeridge DL, et al. Temporal Changes in Pediatric Gastroenteritis after Rotavirus Vaccination in Quebec. *Pediatr Infect Dis J.* 2016;35(5):555–60.
 3. Karafillakis E, Hassounah S, Atchison C. Effectiveness and impact of rotavirus vaccines in Europe, 2006–2014. *Vaccine.* 2015;33(18):2097–107.
 4. Leshem E, Moritz RE, Curns AT, Zhou F, Tate JE, Lopman BA, et al. Rotavirus Vaccines and Health Care Utilization for Diarrhea in the United States (2007–2011). *Pediatrics.* 2014;134(1):15–23.
 5. Gurgel RQ, Illozue C, Correia JB, Centenari C, Oliveira SMT, Cuevas LE. Impact of rotavirus vaccination on diarrhoea mortality and hospital admissions in Brazil. *Trop Med Int Heal.* 2011;16(9):1180–4.
 6. Kumazaki M, Usuku S. Norovirus genotype distribution in outbreaks of acute gastroenteritis among children and older people: An 8-year study. *BMC Infect Dis.* 2016;16(1):1–8.
 7. Bucardo F, Reyes Y, Svensson L, Nordgren J. Predominance of norovirus and sapovirus in nicaragua after implementation of universal rotavirus vaccination. *PLoS One.* 2014;9(5):1–8.
 8. McAtee CL, Webman R, Gilman RH, Mejia C, Bern C, Apaza S, et al. Burden of norovirus and rotavirus in children after rotavirus vaccine introduction, Cochabamba, Bolivia. *Am J Trop Med Hyg.* 2016;94(1):212–7.
 9. Payne DC, Vinjé J, Szilagyi PG, Edwards KM, Staat MA, Weinberg GA, et al. Norovirus and Medically Attended Gastroenteritis in U.S. Children. *N Engl J Med.* 2013;368(12):1121–30.

10. Hemming M, Räsänen S, Huhti L, Paloniemi M, Salminen M, Vesikari T. Major reduction of rotavirus, but not norovirus, gastroenteritis in children seen in hospital after the introduction of RotaTeq vaccine into the National Immunization Programme in Finland. *Eur J Pediatr.* 2013;172(6):739–46.
11. Lopman BA, Steele D, Kirkwood CD, Parashar UD. The Vast and Varied Global Burden of Norovirus: Prospects for Prevention and Control. *PLoS Med.* 2016;13(4):1–12.
12. Sharia M Ahmed, Aron J Hall, Anne E Robinson, Linda Verhoef, Prasanna Premkumar, Umesh D Parashar, Marion Koopmans BAL. Global prevalence of norovirus in cases of gastroenteritis: a systematic review and meta-analysis. *Lancet Infect Dis.* 2014;14:725–30.
13. Mawatari M, Kato Y. Norovirus Gastroenteritis. *Emerg Infect Dis Clin Case Stud.* 2014;203–12.
14. Siebenga JJ, Vennema H, Zheng D, Vinjé J, Lee BE, Pang X, et al. Norovirus Illness Is a Global Problem: Emergence and Spread of Norovirus GII.4 Variants, 2001–2007. *J Infect Dis [Internet].* 2009;200(5):802–12. Available from: <https://academic.oup.com/jid/article-lookup/doi/10.1086/605127>
15. Lanata CF, Fischer-Walker CL, Olascoaga AC, Torres CX, Aryee MJ, Black RE. Global Causes of Diarrheal Disease Mortality in Children <5 Years of Age: A Systematic Review. *PLoS One.* 2013;8(9).
16. Riera-Montes M, O’Ryan M, Verstraeten T. Norovirus and Rotavirus disease severity in children. *Pediatr Infect Dis J.* 2017;37(6):1.
17. Bartsch SM, Lopman BA, Ozawa S, Hall AJ, Lee BY. Global economic burden of norovirus gastroenteritis. *PLoS One.* 2016;11(4):1–16.
18. Cortes-Penfield NW, Ramani S, Estes MK, Atmar RL. Prospects and Challenges in the Development of a Norovirus Vaccine. *Clin Ther.* 2017;39(8):1537–49.

19. Lee I, Kuntz G, Stevenson KB. Guideline for the Prevention and Control of Norovirus Gastroenteritis. *Prevention*. 2013;26(3):1–52.
20. Cochran WG. Some Methods for Strengthening the Common χ^2 Tests. *Biometrics*. 1954;10(4):417–51.
21. Borenstein M, Higgins J, Hedges L, Rothstein H. Basics of meta-analysis: I² is not an absolute measure of heterogeneity. *Res Synth Methods*. 2017;8(1):5–18.
22. Nayak MK, Chatterjee D, Nataraju SM, Pativada M, Mitra U, Chatterjee MK, et al. A new variant of Norovirus GII.4/2007 and inter-genotype recombinant strains of NVGII causing acute watery diarrhoea among children in Kolkata, India. *J Clin Virol*. 2009;45(3):223–9.
23. Ramirez S, Giannanco G, De Grazia S, Colomba C, Martella V, Arista S. Emerging GII.4 Norovirus Variants Affect Children With Diarrhea in Palermo, Italy in 2006. *J Med Virol*. 2009;81:139–45.
24. Cheng W xia, Ye X hua, Yang X mei, Li Y ning, Jin M, Jin Y, et al. Epidemiological study of human calicivirus infection in children with gastroenteritis in Lanzhou from 2001 to 2007. *Arch Virol*. 2010;155(4):553–5.
25. Alam A, Qureshi S, Vinjé J, Zaidi A. Genetic Characterization of Norovirus Strains in Hospitalized Children From Pakistan. *J Med Virol*. 2016;88:216–23.
26. Muhsen K, Kassem E, Rubinstein U, Schachter Y, Kremer A, Goren S, et al. Incidence and characteristics of sporadic norovirus gastroenteritis associated with hospitalization of children less than 5 years of age in Israel. *Pediatr Infect Dis J*. 2013;32(6):688–90.
27. Chaimongkol N, Khamrin P, Malasao R, Thongprachum A, Kongsricharoern T, Ukarapol N, et al. Molecular Characterization of Norovirus Variants and Genetic Diversity of Noroviruses and Sapoviruses in Thailand. *J Med Virol*. 2014;86:1210–8.
28. Kittigul L, Pombubpa K, Taweekate Y, Yeephoo T, Khamrin P, Ushijima H. Molecular

- Characterization of Rotaviruses, Noroviruses, Sapovirus, and Adenoviruses in Patients With Acute Gastroenteritis in Thailand. *J Med Virol.* 2009;81:345–53.
29. Bodhidatta L, Abente E, Neesanant P, Nakjarung K, Sirichote P, Bunyarakyothin G, et al. Molecular Epidemiology and Genotype Distribution of Noroviruses in Children in Thailand From 2004 to 2010: A Multi-Site Study. *J Med Virol.* 2015;87:664–74.
30. Dai Y chun, Hu G fang, Zhang X fu, Song C lei, Xiang W long, Wu X bo, et al. Molecular epidemiology of norovirus gastroenteritis in children in Jiangmen, China, 2005-2007. *Arch Virol.* 2011;156(9):1641–6.
31. Cho HG, Lee SG, Kim JE, Yu KS, Lee DY, Park PH, et al. Molecular epidemiology of norovirus GII.4 variants in children under 5 years with sporadic acute gastroenteritis in South Korea during 2006-2013. *J Clin Virol.* 2014;61(3):340–4.
32. Zhirakovskaya E, Tikunov AY, Bodnev SA, Klemesheva V V., Netesov S V., Tikunova N V. Molecular Epidemiology of Noroviruses Associated With Sporadic Gastroenteritis in Children in Novosibirsk, Russia, 2003–2012. *J Med Virol.* 2015;87:740–53.
33. Santos VS, Gurgel RQ, Cavalcante SMM, Kirby A, Café LP, Souto MJ, et al. Acute norovirus gastroenteritis in children in a highly rotavirus-vaccinated population in Northeast Brazil. *J Clin Virol [Internet].* 2017;88:33–8. Available from: <http://dx.doi.org/10.1016/j.jcv.2016.10.015>
34. Hoa-Tran TN, Nakagomi T, Sano D, Sherchand JB, Pandey BD, Cunliffe NA, et al. Molecular epidemiology of noroviruses detected in Nepalese children with acute diarrhea between 2005 and 2011: Increase and predominance of minor genotype GII.13. *Infect Genet Evol.* 2015;30:27–36.
35. Mans J, Murray TY, Nadan S, Netshikweta R, Page NA, Taylor MB. Norovirus diversity in children with gastroenteritis in South Africa from 2009 to 2013: GII.4 variants and

- recombinant strains predominate. *Epidemiol Infect.* 2016;144(5):907–16.
36. Puustinen L, Blazevic V, Huhti L, Szakal ED, Halkosalo A, Salminen M, et al. Norovirus genotypes in endemic acute gastroenteritis of infants and children in Finland between 1994 and 2007. *Epidemiol Infect.* 2012;140(2):268–75.
 37. Iturriza-Gómara M, Elliot AJ, Dockery C, Fleming DM, Gray JJ. Structured surveillance of infectious intestinal disease in pre-school children in the community: “The Nappy Study.” *Epidemiol Infect.* 2009;137(7):922–31.
 38. Kulkarni R, Patel A, Bhalla S, Chhabra P, Cherian S, Chitambar SD. Characterization of GII.4 noroviruses circulating among children with acute gastroenteritis in Pune, India: 2005–2013. *Infect Genet Evol.* 2016;37:163–73.
 39. Abugalia M, Cuevas L, Kirby A, Dove W, Nakagomi O, Nakagomi T, et al. Clinical Features and Molecular Epidemiology of Rotavirus and Norovirus Infections in Libyan Children. *J Med Virol.* 2011;83:1849–56.
 40. Trang N V, Luan LT, Kim-Anh LT, Hau VTB, Nhung LTH. Detection and Molecular Characterization of Noroviruses and Sapoviruses in Children Admitted to Hospital With Acute Gastroenteritis in Vietnam. *J Med Virol.* 2012;84:290–7.
 41. Al-Rashidi A, Chehaded W, Szucs G, Albert MJ. Different Norovirus Genotypes in Patients With Gastroenteritis in Kuwait. *J Med Virol.* 2013;85:1161–618.
 42. Thongprachum A, Khamrin P, Chan-It W, Malasao R, Chaimongkol N, Okitsu S, et al. Emergence of Norovirus GII/4 2006a and 2006b variants in hospitalized children with acute gastroenteritis in Thailand. *Clin Lab.* 2013;59(3–4):271–6.
 43. Nataraju SM, Ganesh B, Das S, Chowdhury S, Nayak MK, Ghosh M, et al. Emergence of noroviruses homologous to strains reported from Djibouti (horn of Africa), Brazil, Italy, Japan and USA among children in Kolkata, India. *Eur Rev Med Pharmacol Sci.*

- 2010;14(9):789–94.
44. Jin M, Xie H, Duan Z, Liu N, Zhang Q, Wu B, et al. Emergence of the GII4/2006b Variant and Recombinant Noroviruses in China. *J Med Virol*. 2008;80:1997–2004.
 45. Raboni SM, Damasio GAC, Ferreira CE, Pereira LA, Nogueira MB, Vidal LR, et al. Acute gastroenteritis and enteric viruses in hospitalised children in southern Brazil: aetiology, seasonality and clinical outcomes. *Mem Inst Oswaldo Cruz*. 2014 Jul;109(4):428–35.
 46. Melhem NM, Zarake H, Kreidieh K, Ali Z, Hammadi M, Ghanem S, et al. Clinical and epidemiological characteristics of norovirus gastroenteritis among hospitalized children in Lebanon. *World J Gastroenterol*. 2016;22(48):10557–65.
 47. My PVT, Thompson CN, Phuc H Le, Tuyet PTN, Vinh H, Nguyen VMH, et al. Endemic norovirus infections in children, Ho Chi Minh City, Vietnam, 2009–2010. *Emerg Infect Dis*. 2013;19(6):29–32.
 48. Yang SY, Hwang KP, Wu FT, Wu HS, Hsiung CA, Chang WC, et al. Epidemiology and Clinical Peculiarities of Norovirus and Rotavirus Infection in Hospitalized Young Children with Acute Diarrhea in Taiwan, 2009. *J Microbiol Immunol Infect*. 2010;43(6):506–14.
 49. Moyo S, Hanevik K, Blomberg B, Kommedal O, Vainio K, Maselle S, et al. Genetic diversity of norovirus in hospitalised diarrhoeic children and asymptomatic controls in Dar es Salaam, Tanzania. *Infect Genet Evol*. 2014;26:340–7.
 50. Tan D, Deng L, Wang M, Li X, Ma Y, Liu W. High Prevalence and Genetic Diversity of Noroviruses Among Children With Sporadic Acute Gastroenteritis in Nanning City, China, 2010–2011. *J Med Virol*. 2015;87:498–503.
 51. Lu L, Jia R, Zhong H, Xu M, Su L, Cao L, et al. Molecular characterization and multiple infections of rotavirus, norovirus, sapovirus, astrovirus and adenovirus in outpatients with sporadic gastroenteritis in Shanghai, China, 2010–2011. *Arch Virol*. 2015;160(5):1229–

- 38.
52. Fu J-G, Ai J, Zhang J, Wu Q-B, Qi X, Ji H, et al. Molecular Epidemiology of Genogroup II Norovirus Infection Among Hospitalized Children With Acute Gastroenteritis in Suzhou (Jiangsu, China) From 2010 to 2013. *J Med Virol.* 2016;88:954–60.
 53. Rahman M, Rahman R, Nahar S, Hossain S, Ahmed S, Faruque ASG, et al. Norovirus Diarrhea in Bangladesh, 2010–2014: Prevalence, Clinical Features, and Genotypes. *J Med Virol.* 2016;88:1742–50.
 54. Becker-Dreps S, Bucardo F, Vilchez S, Enrique Zambrana L, Liu L, Weber DJ, et al. Etiology of Childhood Diarrhea Following Rotavirus Vaccine Introduction: A Prospective, Population-Based Study in Nicaragua. *Pediatr Infect Dis J.* 2014;33(11):1156–63.
 55. El Qazoui M, Oumzil H, Baassi L, El Omari N, Sadki K, Amzazi S, et al. Rotavirus and norovirus infections among acute gastroenteritis children in Morocco. *BMC Infect Dis.* 2014;14(1):1–9.
 56. Zhang S, Chen T-H, Wang J, Dong C, Pan J, Moe C, et al. Symptomatic and Asymptomatic Infections of Rotavirus, Norovirus, and Adenovirus Among Hospitalized Children in Xi'an, China. *J Med Virol.* 2011;83:1476–84.
 57. Chhabra P, Samoilovich E, Yermalovich M, Chernyshova L, Gheorghita S, Cojocaru R, et al. Viral gastroenteritis in rotavirus negative hospitalized children <5 years of age from the independent states of the former Soviet Union. *Infect Genet Evol.* 2014;28:283–8.
 58. Zhang P, Chen L, Fu Y, Ji L, Wu X, Xu D, et al. Clinical and molecular analyses of norovirus-associated sporadic acute gastroenteritis: The emergence of GII.17 over GII.4, Huzhou, China, 2015. *BMC Infect Dis.* 2016;16(1):1–8.
 59. Trang N V, Choisy M, Nakagomi T, Chinh NTM, Doan YH, Yamashiro T, et al. Determination of cut-off cycle threshold values in routine RT-PCR assays to assist

- differential diagnosis of norovirus in children hospitalized for acute gastroenteritis. *Epidemiol Infect.* 2015;143(15):3292–9.
60. Chen SY, Feng Y, Chao HC, Lai MW, Huang WL, Lin CY, et al. Emergence in Taiwan of novel norovirus GII.4 variants causing acute gastroenteritis and intestinal haemorrhage in children. *J Med Microbiol.* 2015;64(2015):544–50.
 61. Zhang SX, Li L, Yin JW, Jin M, Kong XY, Pang LL, et al. Emergence of human caliciviruses among diarrhea cases in southwest China. *BMC Infect Dis.* 2016;16(1):1–9.
 62. Nasab SDM, Sabahi F, Makvandi M, Samiee SM, Nadji SA, Ravanshad M. Epidemiology of rotavirus-norovirus co-infection and determination of norovirus genogrouping among children with acute gastroenteritis in Tehran, Iran. *Iran Biomed J.* 2016;20(5):280–6.
 63. Dábilla N, Nunes Vieira Almeida T, Carvalho Rebouças Oliveira A, Kipnis A, Neres Silva T, Souza Fiaccadori F, et al. Norovirus in feces and nasopharyngeal swab of children with and without acute gastroenteritis symptoms: First report of GI.5 in Brazil and GI.3 in nasopharyngeal swab. *J Clin Virol.* 2017;87:60–6.
 64. Chan MCW, Leung TF, Kwok AK, Lee N, Chan PKS. Characteristics of patients infected with norovirus GII.4 Sydney 2012, Hong Kong, China. *Emerg Infect Dis [Internet].* 2014 Apr [cited 2016 Apr 3];20(4):658–61. Available from: <http://www.scopus.com/inward/record.url?eid=2-s2.0-84922067029&partnerID=tZOTx3y1>
 65. Chen SY, Tsai CN, Chen CL, Chao HC, Lee YS, Lai MW, et al. Severe viral gastroenteritis in children after suboptimal rotavirus immunization in Taiwan. *Pediatr Infect Dis J.* 2013;32(12):1335–9.
 66. Desai R, Hembree CD, Handel A, Matthews JE, Dickey BW, McDonald S, et al. Severe outcomes are associated with genogroup 2 genotype 4 norovirus outbreaks: A systematic literature review. *Clin Infect Dis.* 2012;55(2):189–93.

67. Siebenga JJ, Vennema H, Renckens B, de Bruin E, van der Veer B, Siezen RJ, et al.
Epochal Evolution of GGII.4 Norovirus Capsid Proteins from 1995 to 2006. *J Virol.*
2007;81(18):9932–41.

Supporting Information

S1 Table. Global Norovirus circulating genotypes among children <5 years old.

Supplementary Appendix. Search Strategy.

Fig 1. Flow diagram of study selection.

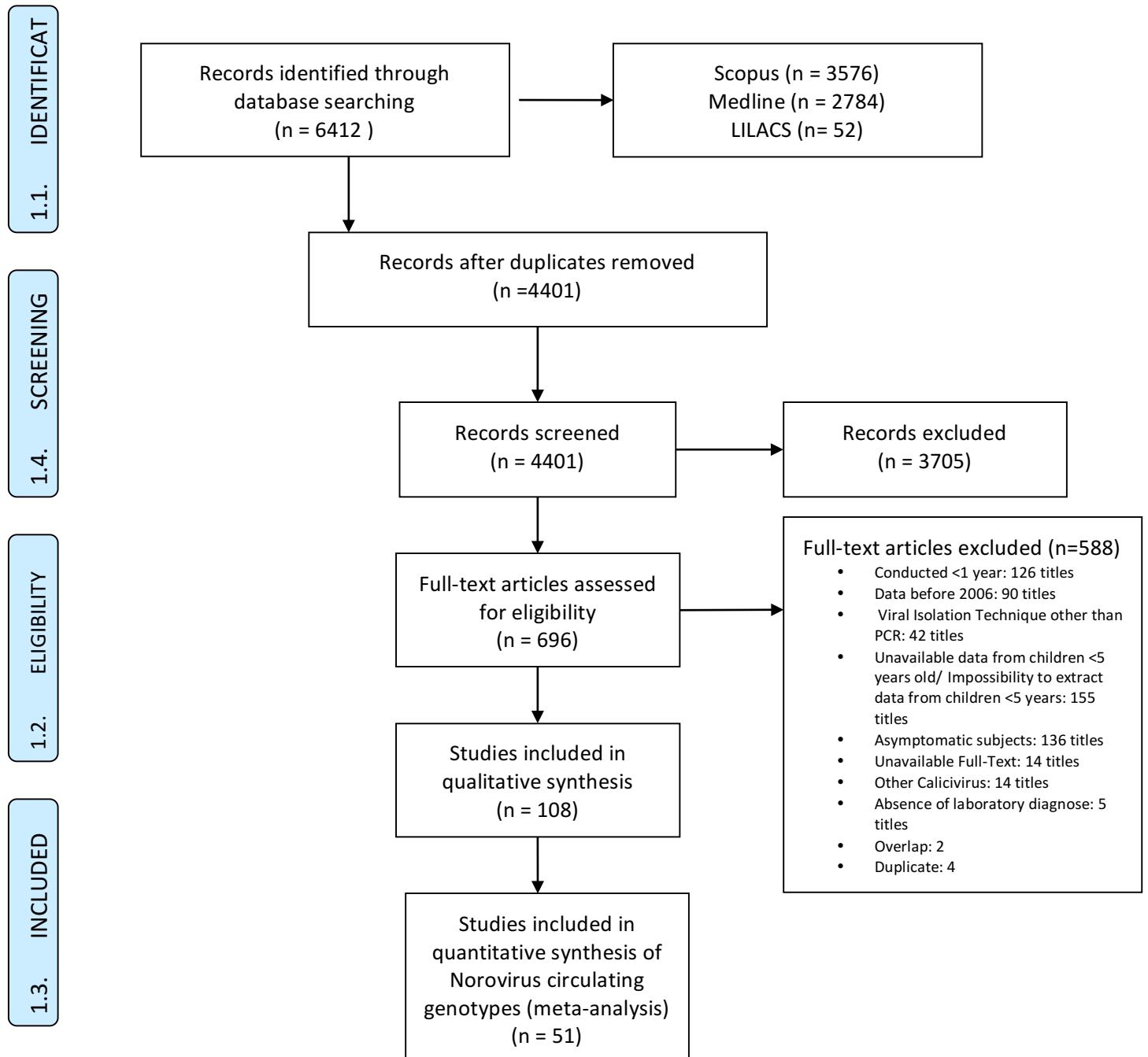


Table 1. Norovirus specimens by genogroups from 2006 to 2015.

Region	GI			GII		
	2006-2008	2009-2011	2012-2015	2006-2008	2009-2011	2012-2015
America	10.8% (6.1–18.2)	3.4% (0.7–14.7)	6.5% (0.4–56.6)	89.2% (81.8–93.4)	96.6% (85.3–99.3)	93.5% (43.4–99.6)
Africa	7.0% (3.7–12.9)	4.4% (2.0–9.4)	7.3% (4.0–12.2)	93.0% (87.1–96.3)	95.6% (91.2–98.0)	92.7% (87.8–96.0)
Asia	4.7% (2.7–7.9)	1.7% (0.7–3.9)	2.8% (1.2–6.0)	95.3% (92.1–97.3)	98.3% (96.1–99.3)	97.3% (94.0–98.8)
Europe	1.8% (0.5–6.8)	5.1% (2.0–12.3)	-	98.2% (93.2–99.5)	94.9% (87.7–98.0)	-

Fig 2. Norovirus circulating genotypes from 2006 to 2008.

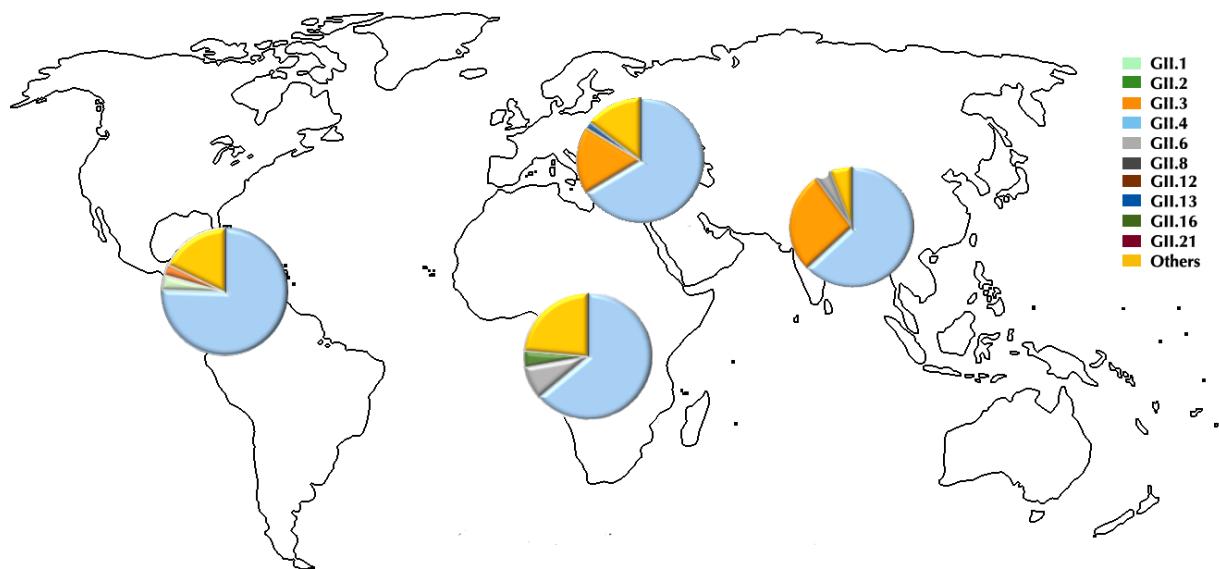


Fig 3. Norovirus circulating genotypes from 2009 to 2011.

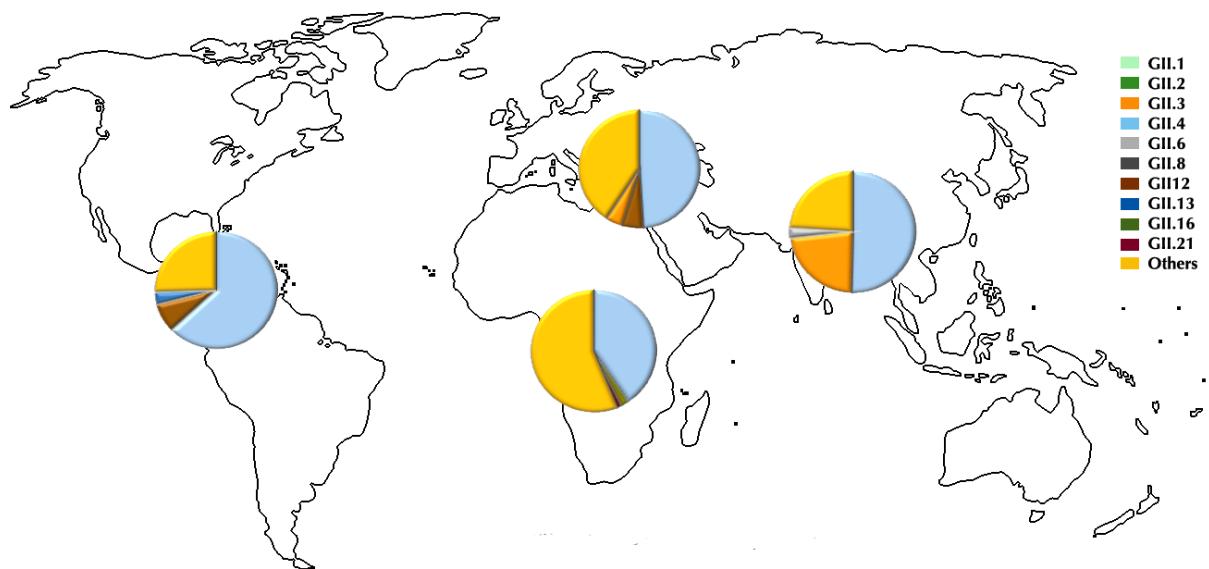


Fig 4. Norovirus circulating genotypes from 2012 to 2015.

