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ATTACHED BIOMASS GROWTH AND SUBSTRATE UTILIZATION RATE IN A MOVING BED BIOFILM REACTOR

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Abstract - A moving bed bioreactor containing cubes of polyether foam immersed in a synthetic wastewater (an aqueous mixture of meat extract, yeast extract, dextrose, meat peptone, ammonium chloride, potassium chloride, sodium chloride, sodium bicarbonate, potassium mono-hydrogen-phosphate and magnesium sulphate) was used to evaluate bacterial growth and biomass yield parameters based on Monod's equation. The wastewater was supplied in the bottom of the equipment flowing ascending in parallel with a diffused air current that provided the mixing of the reactor content. Suspended and attached biomass concentration was measured through gravimetric methods. Good agreement was found between experimental kinetic parameters values and those obtained by other researchers. The only significant difference was the high global biomass content about 2 times the values obtained in conventional processes, providing high performance with volumetric loading rates up to $5.5 \text{ kg COD/m}^3/d$.

Keywords: Fluidized bed; Immobilized biomass; Hybrid reactors.

INTRODUCTION

Biological wastewater treatment has been performed in many different ways since the beginning of the 20th Century (Tyagi and Vembu, 1990). After the registration of the activated sludge patent in England in 1914, some modifications of this process were conceived, including the branch of moving bed biofilm reactors that operate with high biomass concentrations, providing high substrate removal rates (Heijnem et al., 1993).

In 1970's, in the USA, the ECOLOTROL[®] company patented another technique of wastewater treatment using biomass attached on inert carrier particles, giving arise to several moving bed biofilm reactors (Sutton e Mishra, 1994). Fan et al. (1989) have shown some systems involving the contact between solid, liquid and gas phases in a single equipment with special emphasis on air-lift, turbulent bed and fluidized bed reactors (Ouyang e Liaw, 1994; Lazarova e Manem, 1994; Tijhuis et al.,

1994). Recently, Sok'ol and Korpal (2006) used an inverse fluidized bed biofilm reactor (IFBBR) to treat a synthetic wastewater containing phenols. The most significant differences among these equipments are due to geometric design and operating strategy.

The particles of inert carriers have a capital role in this set of processes. According to Tavares et al. (1995), the microorganisms produce a kind of natural polymer responsible for their adhesion on the inert carrier surface developing the biofilm. Table 1 lists typical inert carriers reported by other authors.

The study consisted of biodegradation kinetics, suspended and attached biomass contents and organic matter removal rate - very important issues for reactor design. Most of the mathematical models presented in the literature are based on Monod's equation (Tchobanoglous et al., 2003), involving an endogenous decay term, as follows:

$$\mathbf{r'_g} = \frac{\mu_m X S}{K_s + S} - k_d X \tag{1}$$

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Inert Carrier	Diameter (mm)	Density	(% v/v)	Reference
Sand	0.20	2.65	10	Heijnem et al. (1993); Dirkzwager et al. (1993)
Basalt	0.26	3.00	5	Heijnem et al. (1993); Tijhuis et al. (1994)
Granulated activated carbon	0.40-0.59	1.50	NA	Coelhoso et al. (1992); Ouyang and Liaw (1994)
Hollow polyethylene cylinders	10	0.95	5	Rusten et al. (1994); Sok'ol and Korpal (2006)
Particles of polymers	2.7	1.18	NA	Lazarova and Manen (1994)

Table 1: Some inert carriers reported in the literature

NA - no data available.

Laspidou (2003) presents a more complete kinetic model, taking in account seven chemical (carbonaceous, nitrogenous and phosphorous) species involved in the biodegradation reaction, but it requires further experiments that are beyond the scope of this work.

Literature data about the use of polyether foam as inert carriers for wastewater treatment are rare. Tharp and Frymier (1986); Tyagi and Vembu (1990); Gilligan and Morper (1999) report some results using polyurethane foam pads as support for biomass fixation related to the patented processes CAPTOR and LINPOR, that exhibit some operational differences in relation to the reactor used in this study, but are useful to make a comparison of their performance.

MATERIAL AND METHODS

The study was developed using a 3.8 liter moving bed biofilm reactor whose characteristics are listed below. The wastewater was synthesized by mixing meat extract, yeast extract, dextrose, meat peptone, ammonium chloride, potassium chloride, sodium chloride, potassium chloride, sodium bicarbonate, potassium mono-hydrogen-phosphate and magnesium sulphate, adapted from Hirata (2000). The ratio C:N:P was set around 100:5:1, following Eckenfelder's biodegradation criteria (Eckenfelder, 1992).

The determination of kinetic parameters and suspended biomass content was based on soluble and total COD measurements according to Tchobanoglous et al. (2003). A liquid sample containing suspended biomass was collected from the top of the reactor and its COD was measured in duplicate in two different ways: in the whole sample - whose COD value quantified the soluble substrate plus the suspended biomass content, and in a filtered sample aliquot to evaluate the COD due to the soluble substrate.

The attached biomass was measured gravimetrically through the weight difference between bare and covered samples of the carrier, following the standard method applied to the determination of total suspended solids (Clesceri et al., 1998).

Reactor Characteristics:

Internal diameter	0.60	m
Cylindrical body diameter	0.75	m
Conical adapter height	0.10	m
Top diameter	0.12	m
Cylindrical top height	0.08	m
Free board	0.10	m

The inert carrier used was polyether foam cubes measuring 0.005 m on each side. This material has a density of 0.65, hence is lighter than water, and a porosity of about 95 %.

A set of experimental situations was selected to cover a representative range of organic load and hydraulic retention time, as shown in Table 2.

The experiments were carried out using the maximum content of polyether foam (0.13% by volume) and the air flow of 2 L per minute. The optimal content was obtained using hydrodynamic tests, according to Marques (2003), corresponding to the maximum concentration of solids in which a stable operation could be achieved without any bed clogging. This percentage of support represents only the volume fraction occupied by the solid framework of foam pads. If the void space is included, the support content increases to 2.0 percent.

Table 2: Experimental parameters and respective ranges of studied values

Parameter	Dimension	Values
Polyether foam fraction	% v/v	0.13 (2.70 g)
Air flowrate	L/min	2.0
Hydraulic retention time	h	2; 3; 4; 5
COD level	mg/L	400; 500; 600; 1000

RESULTS AND DISCUSSION

The experimental results obtained from several runs are listed in Table 3.

The kinetic parameters of Monod's model were obtained through data regression, by taking into account endogenous decay as discussed before. The optimized values found are listed in Table 4.

Biomass Attachment Capacity of Polyether Foam

The assessment of biomass fixation in polyether foam cubes (that include the biomass occluded in macroporous media) was carried out through typical biodegradation tests by measuring gravimetrically the increase of fixed biomass content with varying operation time. The results are shown in Figure 1.

It is clear that biomass fixation in polyether foam is very fast in comparison with the colonization of carbon particles reported by Tyagi and Vembu (1990). This process takes several days to be completed because the fixation of biomass on particle surface depends on the generation of a polysaccharide by the microorganisms, according to Laspidou (2003). Probably, in the polyether matrix, the main mechanism involved was the entrapment of biomass flocks instead of the attachment of microorganisms on the surface. Although the biomass can be loose in the three-dimensional frame of the inert carrier, high reactor biomass contents are attained. After about 30 hours of continuous operation, the system becomes saturated.

Tabl	e 3:	Ex	perimental	results	under	investigated	operational	conditions

Run	Carrier	Q (mL/min)	S _o (mg/L)	$\theta = \theta_{c} (d)$	S (mg/L)	X (mg/L)
1	-	25.2	534	0.106	116	138
2	-	16.8	1003	0.158	100	452
3	-	21.0	632	0.127	179	91
4	-	21.0	569	0.127	340	76
5	-	21.0	350	0.127	69	141
6	POL	25.2	460	0.106	50	166
7	POL	25.2	397	0.106	23	23
8	POL	31.5	400	0.084	124	53
9	POL	16.8	334	0.158	39	39
10	POL	16.8	256	0.158	48	12
11	POL	21.0	417	0.127	108	13
12	POL	16.8	967	0.158	98	94
13	POL	21.0	484	0.127	150	50
14	POL	12.6	589	0.211	210	56

Note: The polyether foam content (POL) was fixed equal to 0.13% v/v and the hydraulic retention time (θ) was established equal to the cells age (θ_c) because the reactor was operated without recycle.

Table 4: Optimized values of kinetic and biomass growth parameters

Parameter	Unit	Value
μ _m	d ⁻¹	3
Ks	mg COD/L	74
k _d	d ⁻¹	0.07
Y	mg TSS/mg COD	0.44
k	d ⁻¹	4.1



Figure 1: Biomass to carrier mass ratio versus time; Q=25.2 mL/min; COD=397 mg/L; Qar=2 L/min

Effect of Organic Load on The Substrate Removal

The activated sludge reactors show a good performance at relatively low organic load (Tchobanoglous et al., 2003). Beyond a certain limit their efficiencies tend to decay as shown in Figure 2.

The substrate utilization rate increases with the organic load until a given limit. The optimal values achieved were at 3.3 kg $COD/m^3/d$, corresponding to a substrate removal rate of 62 kg COD/kg biomass/d. Thus, the moving bed bioreactor used provided a substrate removal capacity about 2 times the values obtained in a conventional activated sludge reactor

using real sanitary effluent (Tchobanoglous et al., 2003), just as predicted by Heijnem et al. (1993). Of course, it must be taken into account that the wastewater used in all the experiments was synthesized and probably more easily degradable than real sanitary wastewater.

Figure 3 shows another comparison involving the organic load (OL) and the volumetric load rate (VLR). There is a linear relation between these two parameters, just like that reported by Jianlong et al. (2000).

These results can be compared with previously obtained data using similar polymeric materials as biomass support, as shown in Table 5.



Figure 2: Correlation between substrate utilization rate and volumetric load rate



Figure 3: Correlation between organic load and volumetric substrate removal rate

T	abl	e	5:	Com	pariso	n betwee	en this	work	and and	lit	erature o	lata
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Process	Support Type	Support Content (%)	Biomass Content (mg/L)	Reference
CAPTOR®	Polyurethane foam pads	15 - 75	7,000 - 9,000	Tyagi and Vembu (1990); Tharper and Frymier (1986)
LINPOR®	Polyurethane foam pads	10 - 40	5,000 - 8,000	Tyagi and Vembu (1990); Gilligan and Morper (1999)
This work	Polyether foam pads	2	650	-
This work	Polyether foam pads	10 - 40	$3,250 - 13,000^{(*)}$	-

(*)Extrapolation based on LINPOR process support content range.

Although the support and biomass contents found in this study are out of the range obtained by the other researchers, if a linear extrapolation in biomass content, based on LINPOR[®] process support content, is assumed, the estimated values of biomass concentration would vary from 3,250 to 13,000 mg/L. For instance, operational limitations such as clogging problems, common in reactors with reduced internal diameter, were not observed.

CONCLUSIONS

The kinetic parameter values obtained are not significantly different from those reported in the literature. The organic load and the substrate utilization rates achieved were so high due to the high biomass to carrier ratios, whose maximum value was about 0.8 kg biomass/kg inert carrier.

This result confirms that moving bed biofilm reactors tolerate around 2 times the volumetric organic loads experienced by the other modalities of activated sludge reactor processes.

The main advantage of this kind of process is the possibility of building a wastewater treatment plant in small areas. Another important issue is that a conventional plant can have its capacity increased by using suspended inert carriers like polyether or polyurethane foam pads.

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NOMENCLATURE

COD	chemical oxygen demand	mg/L
k _d	endogenous decay	h^{-1}
	coefficient	
Ks	half-velocity constant	mg/L
OL	organic load	kgCOD/d
Q	feed flowrate	mL/min
Q _{ar}	air flowrate	L/min
R	biomass to carrier mass ratio	
r _g	net rate of bacterial growth	mass/volume/
0		time
S	concentration of growth-	mg/L
	limiting substrate in the	

	reactor content	
So	concentration of growth-	mg/L
	limiting substrate in the feed	
	stream	
U	volumetric substrate	kg COD/kg
	removal rate	biomass/d
VLR	volumetric load rate	kgCOD/m ³ /d
Х	biomass concentration	mg/L
μ	specific growth rate	h^{-1}
$\mu_{\rm m}$	maximum specific growth	h^{-1}
• •••	rate	
θ	hydraulic retention time	h
$\theta_{\rm c}$	mean cell retention time or	h.
-	sludge age	

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